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(54) Title: MATERIALS AND METHODS FOR GENE THERAPY <div style="display: flex; align-items: center;"> <div style="margin-right: 20px;"> <p>A-AT</p> <p>B-AT</p> <p>C-AT</p> <p>E-AT</p> </div> <div> <p style="text-align: center;">U1a</p> <p style="text-align: center;">U1b</p> <p style="text-align: center;">CMV</p> <p style="text-align: center;">ITR ELF hAAT An Tk neo An ITR</p> </div> </div>			
(57) Abstract The subject invention concerns materials and methods for gene therapy. One aspect of the invention pertains to vectors which can be used to effect genetic therapy in animals or humans having genetic disorders where expression of high levels of a protein of interest are required to treat or correct the disorder. The subject invention also pertains to methods for treating animals or humans in need of gene therapy to treat or correct a genetic disorder. The materials and methods of the invention can be used to provide therapeutically effective levels of a protein that is non-functional, or that is absent or deficient in the animal or human to be treated. In one embodiment, the materials and methods can be used to treat alpha-1-antitrypsin deficiency.			

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DESCRIPTIONMATERIALS AND METHODS FOR GENE THERAPY

5 The subject invention was made with government support under a research project supported by National Institute of Health NHLBI Grant No. HL 59412. The government has certain rights in this invention.

Cross-Reference to a Related Application

10 This application claims priority from provisional application U.S. Serial No. 60/083,025, filed April 24, 1998.

Background of the Invention

15 Alpha-1-antitrypsin (AAT) deficiency is the second most common monogenic lung disease in man, accounting for approximately 3% of all early deaths due to obstructive pulmonary disease. AAT protein is normally produced in the liver, secreted into the serum and circulated to the lung where it protects the fine supporting network of elastin fibers from degradation by neutrophil elastase. Current therapy for AAT deficiency includes avoidance of cigarette smoke exposure and weekly intravenous
20 infusions of recombinant human AAT (hAAT) protein. Attempts to devise gene therapy strategies to replace AAT either in the lung itself or within any of a number of other tissues which are capable of AAT secretion have been limited by the short duration of expression from some vectors and by the relatively high circulating levels of AAT which is required for therapeutic effect. Methods of gene therapy have been described in U.S.
25 Patent No. 5,399,346.

 It has recently been demonstrated that adeno-associated virus (AAV) vectors are capable of stable *in vivo* expression and may be less immunogenic than other viral vectors (Flotte *et al.*, 1996; Xiao *et al.*, 1996; Kessler *et al.*, 1996; Jooss *et al.*, 1998). AAV is a non-pathogenic human parvovirus whose life cycle naturally includes a
30 mechanism for long-term latency. In the case of wild-type AAV (wtAAV), this persistence is due to site-specific integration into a site on human chromosome 19 (the AAVSI site) in the majority of cells (Kotin *et al.*, 1990), whereas with recombinant

AAV (rAAV) vectors, persistence appears to be due to a combination of episomal persistence and integration into non-chromosome 19 locations (Afione *et al.*, 1996; Kearns *et al.*, 1996). Recombinant AAV latency also differs from that of wtAAV in that wtAAV is rapidly converted to double-stranded DNA in the absence of helper virus (e.g., adenovirus) infection, while with rAAV leading strand synthesis is delayed in the absence of helper virus (Fisher *et al.*, 1996; Ferrari *et al.*, 1996). U.S. Patent No. 5,658,785 describes adeno-associated virus vectors and methods for gene transfer to cells.

Kessler *et al.* (1996) demonstrated that murine skeletal myofibers transduced by an rAAV vector were capable of sustained secretion of biologically active human erythropoietin (hEpo), apparently without eliciting a significant immune response against the secreted hEpo. See also U.S. Patent No. 5,858,351 issued to Podsakoff *et al.* Likewise, Murphy *et al.* (1997) have observed the expression and secretion of sustained levels of leptin in *ob/ob* mice after AAV muscle transduction. Brantly *et al.* (U.S. Patent No. 5,439,824) disclose methods for increasing expression of AAT using vectors comprising intron II of the human AAT gene. However, the level of leptin expression observed was only in the range of 2 to 5 ng/ml. Therapy for AAT deficiency requires serum levels of at least about 800 μ g/ml. Thus, there remains a need in the art for a means of providing therapeutically beneficial levels of a protein to a person in need of such treatment.

Brief Summary of the Invention

The subject invention concerns materials and methods for gene therapy. One aspect of the invention pertains to vectors which can be used to provide genetic therapy in animals or humans having a genetic disorder where relatively high levels of expression of a protein is required to treat the disorder. The vectors of the invention are based on adeno-associated virus (AAV). The vectors are designed to provide high levels of expression of heterologous DNA contained in the vector. In one embodiment, the vectors comprise AAV inverted terminal repeat sequences and constitutive or regulatable promoters for driving high levels of gene expression. The subject invention also pertains to methods for treating animals or humans in need of gene therapy, e.g., to correct a genetic deficiency disorder.

Brief Description of the Drawings

Figure 1 shows rAAV-AAT vector cassettes used according to the subject invention. The A-AT and B-AT constructs contain the promoters from the small nuclear RNA genes, U1a and U1b, respectively. The C-AT construct contains the CMV promoter, whereas the E-AT vector uses the human elongation factor 1- α (ELF in the figure) promoter. ITR refers to AAV inverted terminal repeat; An refers to polyA signal; Tk refers to the HSV thymidine kinase promoter; *neo* refers to the Tn5 neomycin phosphotransferase gene.

Figure 2 shows hAAT secretion rates *in vitro* from transiently transfected murine C2C12 myoblast cell line using expression vectors according to the subject invention. C-AT does not differ significantly from E-AT, but both differ from A-AT and B-AT ($p < 0.05$) AAT expression was detected using an ELISA assay specific for human AAT.

Figure 3 shows hAAT secretion rates *in vitro* from stably transduced murine C2C12 myoblast cell line using viral particles comprising expression vectors according to the subject invention. The mean rates of secretion from G418-resistant cultures 1 mo after transduction with either packaged E-AT vector or packaged C-AT vector are shown. In each instance, a "low" multiplicity transduction (4×10^5 particles/cell) and a high multiplicity transduction (4×10^6 particles/cell) were performed. E-AT "low" and "high" are greater than "high" multiplicity C-AT ($P = 0.02$) but are not significantly different from each other ($n = 3$). AAT expression was detected using an ELISA assay specific for human AAT.

Figure 4 shows additional constructs tested for hAAT expression. The murine myoblast C2C12 cells were grown in 35-mm wells with approximately 4×10^5 cell per well and were transfected with 5 μ g of the appropriate plasmid DNA using Superfect transfection (Qiagen Inc., CA). Secretion of hAAT into the medium was assessed at 2 days after transfection using an antigen-capture ELISA. Each bar represents the mean of results from three experiments (triplicate in each experiment).

Data from transfection experiments indicate that the expression from p43CB-AT was at least three times higher than that from C-AT *in vitro*.

Figures 5A and 5B show sustained secretion of therapeutic levels of hAAT using either the C-AT vector or the E-AT vector in either SCID or C57BL mice. Figure 5A shows the mean total serum levels of hAAT observed in groups of either SCID (squares)

or C57BL (circles) mice receiving either low dose (5×10^{11} particles) (open symbols) or high dose (1.4×10^{13} particles) (filled symbols) single injections into muscle of the C-AT vector measured at time points ranging from 1 to 16 wk after injection. For each strain, the high-dose curve is significantly different from the low-dose curve ($P=0.009$ for SCID, $P=0.02$ for C57BL), but the strains do not differ from each other. Figure 5B shows analogous data with the E-AT vector. None of these differences were significant.

Figure 5C shows long term secretion of hAAT from murine muscle transduced with C-AT. C57B1/6 or C57B1/6-SCID mice received 3.5×10^{10} IU, 1.4×10^{13} particles/mouse. One year after injection, serum hAAT levels were still 400 $\mu\text{g/ml}$ in C57B1/6-SCID and 200 $\mu\text{g/ml}$ in C57B1/6. This level are comparable with the peak levels observed (800 or 400 $\mu\text{g/ml}$, respectively).

Figure 6 shows an immunoblot of sera taken from several of the C-AT vector-treated mice at 11 weeks after vector administration. Ten microliters of a 1:100 dilution of serum was electrophoresed by 10% SDS/PAGE, blotted, and incubated with 1:1,500 dilution of goat anti-hAAT-horseradish peroxidase conjugate (Cappel/ICN). Samples from three high-dose SCID (h1-h3), one high-dose C57BL (h3), and three low-dose C57BL (lo1-lo3) were included, along with one negative control (saline-injected = sal) serum to indicate the level of reactivity with endogenous mAAT. As a standard, hAAT was added either to negative-control C57BL serum (first hAAT lane) or to PBS (second hAAT) lane to final equivalent serum concentration of 100 $\mu\text{g/ml}$.

Figures 7A and 7B show that some BALB/c mice mount humoral immune responses to hAAT, which correlate with lower serum levels but no observable toxicity. Figure 7A shows serum hAAT levels and Figure 7B shows serum anti-hAAT antibody levels as determined by ELISA performed on serum taken from mice injected with 1×10^{11} particles of the C-AT vector. Each set of symbols represents an individual animal (\square , no. 1; Δ , no. 2; \circ , no. 3). Note the inverse correlation between the presence of antibody and the presence of circulating hAAT.

Figure 8 shows the persistence of rAAV-AAT vector DNA in high molecular weight form. PCR products were amplified from DNA prepared by Hirt extraction from three SCID mice injected 16 wk earlier with 5×10^{11} resistant-particles of C-AT and analyzed by Southern blot. The high molecular weight Hirt pellet (genomic DNA lanes) and the low molecular weight supernatant (episomal DNA lanes) were analyzed

separately. Control lanes include a sample in which an hAAT cDNA plasmid was the template DNA (+) and a control in which water was the template (-). In this internal PCR reaction, a 500-bp product is expected regardless of whether or not the vector genome is integrated.

5 **Figure 9** shows serum hAAT in C57B1/6 mice transduced with C-AT and p43CB-AT. C57B1/6 mice were injected in muscle with C-AT (3.5×10^{10} IU/mouse, 1×10^{12} particles/mouse) or p43CB-AT (6×10^9 IU, 1×10^{12} particles/mouse). The level of hAAT from p43CB-AT were projected based on an estimation of the equivalent dosage (infectious unit) of C-AT.

10 **Figure 10** shows enhancement of CMV promoter activity by a synthetic enhancer in C2C12 cells. The murine myoblast C2C12 cells were grown in 35-mm wells with approximately 4×10^5 cell per well and were transfected with 5 μ g of p.43rmsENC-AT vector DNA using SUPERFECT transfection (Qiagen Inc, CA). Secretion of hAAT into the medium was assessed at 2 days after transfection using an antigen-capture ELISA.

15 Each bar represents the mean of results from one experiment (triplicate).

Figure 11 shows secretion of hAAT from mouse liver cells (HO15) transfected with different constructs. The murine liver cells (HO15) were grown in 35-mm wells with approximately 4×10^5 cell per well and were transfected with 5 μ g of the plasmid DNA using LIPOFECTAMINE reagents (Life Technologies Inc, MD). Secretion of

20 hAAT into the medium was assessed at 2 days after transfection using an antigen-capture ELISA. Each bar represents the mean of results from two experiments (triplicate).

Figure 12 shows secretion of hAAT from mouse liver cells (HO15) transfected using different methods. The murine liver cells (HO15) were grown in 35-mm wells with approximately 4×10^5 cell per well and were transfected with 5 μ g of the p43CB-AT vector

25 using Superfect (Qiagen Inc., CA), FuGENE (Boehringer Mannheim Co, IN), Lipofectin, LipofectAMINE (Life Technologies Inc, MD) reagents and Calcium phosphate (CA-PO₄) transfection. Secretion of hAAT into the medium was assessed at 2 days after transfection using an antigen-capture ELISA. Each bar represents the mean of results from one experiment (triplicate).

30 **Figure 13** shows hAAT secretion from mouse liver transduced with rAAV. C57B1/6 mice were injected with either p43CB-AT, C-AT or E-AT vector either by portal vein or tail vein injection. PV=portal vein injection. TV=tail vein injection.

Figure 14 shows serum hAAT levels in C57Bl/6 mice after intratracheal (IT) injection of C-AT or p43CB-AT vector. Mice received either 10^9 IU of C-AT (open circles), 10^9 IU of p43CB-AT (open triangles) or 10^{10} IU of p43CB-AT (open squares).

Figure 15 shows a map and nucleotide sequence for the vector of the present invention designated as C-AT.

Figure 16 shows a map and nucleotide sequence for the vector of the present invention designated as E-AT.

Figure 17 shows a map and nucleotide sequence for the vector of the present invention designated as dE-AT.

Figure 18 shows a map and nucleotide sequence for the vector of the present invention designated as p43C-AT.

Figure 19 shows a map and nucleotide sequence for the vector of the present invention designated as p43C-AT-IN. This vector includes intron II from human AAT gene to enhance transcription.

Figure 20 shows a map and nucleotide sequence for the vector of the present invention designated as p43CB-AT.

Figure 21 shows a map and nucleotide sequence for the vector of the present invention designated as C-AT2.

Figure 22 shows a map and nucleotide sequence for the vector of the present invention designated as p43msENC-AT. This vector is similar to p43C-AT but also comprises an enhancer sequence upstream of the CMV promoter.

Figure 23 shows a map and nucleotide sequence for the vector of the present invention designated as p43rmsENC-AT. This vector is the same as the p43msENC-AT vector except that the enhancer sequence is in an opposite orientation.

Figure 24 shows a map and nucleotide sequence for the vector of the present invention designated as p43msENCB-AT. This vector is similar to p43CB-AT but also comprises an enhancer sequence upstream of the CMV promoter.

Figure 25 shows a map and nucleotide sequence for the vector of the present invention designated as p43rmsENCB-AT. This vector is the same as p43msENCB-AT except that the enhancer sequence is in an opposite orientation.

Detailed Disclosure of the Invention

The subject invention pertains to novel materials and methods for providing gene therapy to a mammal or human having a condition or disorder, such as genetic deficiency disorders, where high levels of expression of a protein are required to treat the disorder or condition. In one method of the subject invention, a viral vector is introduced into cells of an animal wherein a therapeutic protein is produced, thereby providing genetic therapy for the animal. In one embodiment, a method of the invention comprises introducing into an animal cell or tissue an effective amount of viral particles or vector comprising a recombinant genome which includes heterologous polynucleotide encoding a protein useful in genetic therapy and that can be expressed by the cell or tissue. Expression of the heterologous polynucleotide results in production of the protein. Preferably, the therapeutic protein encoded by the heterologous polynucleotide is a serum protein. In a preferred embodiment, vector material comprising the heterologous polynucleotide is integrated into a chromosome of the cell of the host animal.

In one embodiment, a recombinant polynucleotide vector of the present invention is derived from adeno-associated virus (AAV) and comprises a constitutive or regulatable promoter capable of driving sufficient levels of expression of the heterologous DNA in the viral vector. Preferably, a recombinant vector of the invention comprises inverted terminal repeat sequences of AAV, such as those described in WO 93/24641. In a preferred embodiment, a vector of the present invention comprises polynucleotide sequences of the pTR-UF5 plasmid. The pTR-UF5 plasmid is a modified version of the pTR_{BS}-UF/UF1/UF2/UFB series of plasmids (Zolotukhin *et al.*, 1996). The pTR-UF5 plasmid contains modifications to the sequence encoding the green fluorescent protein (GFP).

Promoters useful with the subject invention include, for example, the cytomegalovirus immediate early promoter (CMV), the human elongation factor 1-alpha promoter (EF1), the small nuclear RNA promoters (U1a and U1b), α -myosin heavy chain promoter, Simian virus 40 promoter (SV40), Rous sarcoma virus promoter (RSV), adenovirus major late promoter, β -actin promoter and hybrid regulatory element comprising a CMV enhancer/ β -actin promoter. These promoters have been shown to be active in a wide range of mammalian cells. In addition to the natural promoters described above, synthetic promoters can be used in the present invention. For example, a synthetic

enhancer randomly assembled from Spc5-12-derived elements including muscle-specific elements, serum response factor binding element (SRE), myocyte-specific enhancer factor-1 (MEF-1), myocyte-specific enhancer factor -2 (MEF-2), transcription enhancer factor-1 (TEF-1) and SP-1 (Li *et al.*, 1999; Deshpande *et al.*, 1997; Stewart *et al.*, 1996; Mitchell *et al.*, 1989; Briggs *et al.*, 1986; Pitluk *et al.*, 1991) can be used in vectors of the invention.

The promoters are operably linked with heterologous DNA encoding the protein of interest. By "operably linked," it is intended that the promoter element is positioned relative to the coding sequence to be capable of effecting expression of the coding sequence.

Promoters particularly useful for expression of a protein in muscle cells include, for example, hybrid CMV/ β -actin promoters, CMV promoters, synthetic promoters and EF1 promoter. Promoters particularly useful for expression of a protein in liver cells include, for example, hybrid CMV/ β -actin promoters and EF1 promoters.

Also contemplated for use with the vectors of the present invention are inducible and cell type specific promoters. For example, Tet-inducible promoters (Clontech, Palo Alto, CA) and VP16-LexA promoters (Nettelbeck *et al.*, 1998) can be used in the present invention.

The vectors can also include introns inserted into the polynucleotide sequence of the vector as a means for increasing expression of heterologous DNA encoding a protein of interest. For example, an intron can be inserted between a promoter sequence and the region coding for the protein of interest on the vector. Introns can also be inserted in the coding regions. Transcriptional enhancer elements which can function to increase levels of transcription from a given promoter can also be included in the vectors of the invention. Enhancers can generally be placed in either orientation, 3' or 5', with respect to promoter sequences.

Heterologous polynucleotide in the recombinant vector can include, for example, polynucleotides encoding normal, functional proteins which provide therapeutic replacement for normal biological function in animals afflicted with genetic disorders which cause the animal to produce a defective protein, or abnormal or deficient levels of that protein. Proteins, and the polynucleotide sequences that encode them, which can be provided by gene therapy using the subject invention include, but are not limited to, anti-

proteases, enzymes, structural proteins, coagulase factors, interleukins, cytokines, growth factors, interferons, and lymphokines. In an exemplified embodiment, heterologous DNA in a recombinant AAV vector encodes human alpha-1-antitrypsin protein.

5 The gene therapy methods of the invention can be performed by *ex vivo* or *in vivo* treatment of the patient's cells or tissues. Cells and tissues contemplated within the scope of the invention include, for example, muscle, liver, lung, skin and other cells and tissues that are capable of producing and secreting serum proteins. The vectors of the invention can be introduced into suitable cells, cell lines or tissue using methods known in the art. The viral particles and vectors can be introduced into cells or tissue *in vitro* or *in vivo*. Methods contemplated include transfection, transduction, injection and inhalation. For example, vectors can be introduced into cells using liposomes containing the subject vectors, by direct transfection with vectors alone, electroporation or by particle bombardment. In an exemplified embodiment, muscle cells are infected *in vivo* by injection of viral particles comprising recombinant vector into muscle tissue of an animal. In another embodiment, liver cells are infected *in vivo* by injection of recombinant virus into either the portal vein or peripheral veins.

15 The methods and materials of the subject invention can be used to provide genetic therapy for any conditions or diseases treatable by protein or cytokine infusion such as, for example, alpha-1-antitrypsin deficiency, hemophilia, adenosine deaminase deficiency, and diabetes. The methods and materials of the subject invention can also be used to provide genetic therapy for treating conditions such as, for example, cancer, autoimmune diseases, neurological disorders, immunodeficiency diseases, and bacterial and viral infections. For example, the present invention can be used to provide genetic therapy to a patient wherein cells from the patient are transformed to express and produce interleukins such as interleukin-2.

20 Animals that can be treated with the materials and methods of the invention include mammals such as bovine, porcine, equine, ovine, feline and canine mammals. Preferably, the mammals are primates such as chimpanzees and humans.

25 The subject invention also concerns cells containing recombinant vectors of the present invention. The cells can be, for example, animal cells such as mammalian cells. Preferably, the cells are human cells. More preferably, the cells are human myofibers or myoblasts, hepatocytes or lung cells. In a preferred embodiment, a recombinant vector

of the present invention is stably integrated into the host cell genome. Cell lines containing the recombinant vectors are also within the scope of the invention.

In an exemplified embodiment, recombinant AAV vectors comprising the human AAT gene (hAAT) using either the CMV promoter (AAV-C-AT) or the human elongation factor 1-alpha (EF1) promoter (AAV-E-AT) to drive expression were constructed and packaged using standard techniques. A murine myoblast cell line, C2C12, was transduced with each vector and expression of hAAT into the medium was measured by ELISA. *In vitro*, the EF1 promoter construct resulted in 10-fold higher hAAT expression than the CMV promoter construct. *In vivo* transduction was performed by injecting doses of up to 1.4×10^{13} Dnase-resistant particles of each vector into skeletal muscles of a number of different strains of mice (including C57B1/6, Balb/c, and SCID). *In vivo*, the CMV promoter construct resulted in higher levels of expression, with sustained serum levels up to 800 $\mu\text{g/ml}$ in SCID mice, approximately 10,000-fold higher than those previously observed with proteins secreted from AAV vectors in muscle. At lower doses in both C57B1/6 and SCID mice, expression was delayed for several weeks, but was sustained for over 10 weeks without declining. Thus, increasing dosage AAV vector via transduction of skeletal muscle provides a means for replacing AAT or other serum proteins.

Transduction of muscle using the vectors of the subject invention presents several advantages in that it is stable, non-toxic, and relatively nonimmunogenic. Furthermore, certain transcription promoters, such as the CMV promoter, which appear to be markedly down-regulated in other contexts have been found to remain active over time as used in the subject invention. Using the materials and methods of the subject invention, microgram/ml serum levels of a therapeutic protein can be achieved. In an exemplified embodiment, the levels of *in vivo* protein expression achieved represent a 10,000-fold or more increase over previously published results. In addition, a dose-effect relationship was demonstrable within the range of doses used, providing for further increases in expression levels as vector dose is increased.

In another embodiment of the invention, recombinant AAV vectors *i.e.*, C-AT, p43C-AT, P43CB-AT, E-AT and dE-AT comprising the human AAT gene (hAAT) using were constructed and packaged using standard techniques. A murine liver cell line, HO15, was transfected with each vector and expression of hAAT into the medium was

measured by ELISA. *In vitro*, transduction with the p43CB-AT vector exhibited the highest level of hAAT expression. *In vivo*, the p43CB-AT vector also gave higher levels of expression. Portal vein administration appeared to be the more efficient route of administration as mice injected in this manner exhibited higher levels of expression than those receiving peripheral vein injections. Transduction of liver offers the same advantages as for muscle, but hepatocytes may be more efficient at secretion of protein.

The dosage of recombinant vector or the virus to be administered to an animal in need of such treatment can be determined by the ordinarily skilled clinician based on various parameters such as mode of administration, duration of treatment, the disease state or condition involved, and the like. Typically, recombinant virus of the invention is administered in doses between 10^5 and 10^{14} infectious units. The recombinant vectors and virus of the present invention can be prepared in formulations using methods and materials known in the art. Numerous formulations can be found in Remington's Pharmaceutical Sciences, 15th Edition (1975).

All publications and patents cited herein are expressly incorporated by reference.

Materials and Methods

Construction of rAAV plasmids. The rAAV-AAT vector plasmids used for these experiments are depicted diagrammatically (Figure 1). Briefly, the plasmid pN2FAT (Garver *et al.* (1987) plasmid was digested with *Xho*I to release 1.8-kb fragment containing the human AAT cDNA along with the SV40 promoter and a polyadenylation signal. This fragment was subcloned into a plasmid, pBlueScript (Stratagene) and, after the removal of the SV40 promoter by *Hind* III digestion and religation, the hAAT cDNA with its polyA signal was released by *Xba*I and *Xho*I digestion. This 1.4-kb *Xba*I-*Xho*I fragment was then cloned in to the pTR-UF5 (an AAV-inverted terminal repeat-containing vector) plasmid (Zolotukhin *et al.*, 1996) between the *Xba*I site 3' to the CMV promoter and the *Xho*I site 5' to the polyoma virus enhancer/HSVthymidine kinase promoter cassette, which drives *neo* in that construct. This yielded the pAAV-CMV-AAT construct (C-AT). Analogous constructs using the promoter from the small nuclear RNA proteins, U1a and U1b, (to give the A-AT and B-AT constructs, respectively) and

human elongation factor 1-alpha (EF1) promoter (to give the E-AT construct) were constructed by substituting each of these promoter cassettes in place of the CMV promoter, between the KpnI and XbaI sites.

The construct, dE-AT derived from E-AT by deletion of the silencer (352 bp) by SAC II-cut (Wakabayashi-Ito *et al.*, 1994). C-AT2 is similar with C-AT except there are SV40 intron and poly (A) sequences flanking the cDNA of hAAT. The p43C-AT was constructed by insertion of hAAT cDNA to an AAV-vector plasmid (p43), which has CMV promoter, intron and poly (A) sequences. The p43CB-AT is derived by replacement of CMV promoter with CMV enhancer and chicken β -actin promoter sequences. The p43C-AT-IN is derived from p43C-AT by insertion of intron II sequences of hAAT gene to hAAT cDNA (Brantly *et al.*, 1995).

Packaging of rAAV vectors. Vectors were packaged using a modification of the method described by Ferrari *et al.* (1997). Briefly, plasmids containing the AAV *rep* and *cap* genes (Li *et al.*, 1997) and the Ad genes (E2a, E4 and VA-RNA) were co-transfected along with the appropriate AAV-AAT vector plasmid into 293 cells grown in Cell Factories (Nunc). Cells were harvested by trypsinization and disrupted by freeze-thaw lysis to release vector virions which were then purified by iodixanol gradient ultracentrifugation followed by heparin sepharose affinity column purification. Alternatively, recombinant virus can be prepared according to methods described in Zolotukhin *et al.* (1999).

Vector preparations had their physical titer assessed by quantitative competitive PCR and their biological titer assessed by infectious center assay. The presence of wild-type AAV was also assessed using these same assays with appropriate internal AAV probes. The high-dose C-AT stock had a particle-titer of 2.0×10^{14} particles/ml and an infectious titer of 5.0×10^{11} infectious units (i.u.)/ml (particle to i.u. ratio = 400:1). The low-dose C-AT measured 8×10^{12} particles/ml and 1.2×10^{10} i.u./ml (particle to i.u. = 667:1). For the E-AT experiments, the titers were 1×10^{13} particles/ml and 2.5×10^{10} i.u./ml (particle to i.u. = 400:1). The low-dose C-AT stock had a wt-like AAV particle titer (*i.e.*, positive AAV genome PCR) equal to 0.1 times the recombinant titer but no detectable infectious wtAAV. The other two preparations had wt-like AAV particle titers $< 10^{-5}$ times the recombinant titer and no detectable infectious wtAAV.

In vitro transfection and transduction experiments. The C2C12 murine myoblast line was used for *in vitro* transfection and transduction experiments. Cells were grown in 35-mm wells with approximately 4×10^5 cells per well and transfected with $5 \mu\text{g}$ of each plasmid DNA using Superfect (Qiagen Corp.). Secretion of hAAT into the medium was assessed at 2 days after transfection using an antigen-capture ELISA assay with standards (Brantly *et al.*, 1991). An SV40 promoter luciferase-expression plasmid, pGL2 (Promega), was used as an internal control. For transduction experiments, cells were grown under similar conditions and were transduced with vector at multiplicities of infection ranging from 4×10^5 to 4×10^6 particles per cell. Cells were then passaged in the presence of geneticin sulfate ($350 \mu\text{g/ml}$) and geneticin-resistant clones were isolated for hAAT secretion studies.

In vivo injection of AAV-C-AT and AAV-E-AT vectors into murine muscle. Mice strains (C57B1/6, SCID, and Balb/c) were obtained from Jackson Laboratories (Bar Harbor, ME) and were handled under specific pathogen-free conditions under a protocol approved by the University of Florida Institutional Animal Care and Use Committee. Animals were anesthetized by metaphane inhalation and aliquots of vector were injected percutaneously into the quadriceps femoris muscles of both hind limbs. The volume of vector ranged from 50 to $100 \mu\text{l}$ per injection site and the total amount of virus injected per animal ranged from 5×10^{10} to 1.4×10^{13} Dnase-resistant particles.

Antigen capture ELISA assay for hAAT expression. Microtiter plates (Immulon 4, Dynex Technologies, Chantilly, VA) were coated with $100 \mu\text{l}$ of a 1:200 dilution of goat anti-human AAT (CAPPEL/ICN) in Vollers buffer ($\text{Na}_2\text{CO}_3=2.76\text{g}$, $\text{NaHCO}_3=1.916\text{g}$, $\text{NaN}_3=0.2\text{g}$, $\text{d.H}_2\text{O}=1$ liter, Adjust PH=9.6) overnight at 4°C . After washing, standards and unknown samples containing hAAT were incubated in the plates at 37°C for 1 hour. After blocking in 3% BSA in PBS-Tween 20 at 37°C for 1 hour, a second antibody (1:1000 dilution of rabbit anti-human AAT, Boehringer Mannheim) was reacted with the captured antigen at 37°C for 1 hour. Detection was performed using a third antibody incubation (1:800 dilution of goat anti-rabbit IgG-peroxidase conjugate, 37°C) followed by *o*-phenylenediamine (OPD, Sigma) detection and measurement of the absorbance at 490nm.

ELISA assay for anti-hAAT and anti-AAV VP3 antibodies. Wells were coated with antigen ($1 \mu\text{g}$ of hAAT or 100ng of VP3) at 4°C overnight, blocked with 3% BSA

and then reacted with dilutions of either test serum or with positive control antibodies at 37°C for 1 hour. After washing, a goat-anti-mouse IgG-peroxidase conjugate was used as a secondary antibody (1:1500 dilution) to detect bound anti-AAT antibody, using a standard OPD reaction, as described above. Antibody levels were quantitated by comparison with a standard curve generated by reacting dilutions of known positive monoclonal antibodies against VP3 and hAAT.

Lymphocyte proliferation assays to detect cell-mediated immune responses.

Lymphocyte proliferation assays were performed in order to detect T cell responses to the hAAT and VP3 antigens. Freshly isolated splenocytes were grown in primary culture in 96 well plates coated with 0, 0.1, 1, and 10 µg of either hAAT or VP3 in RPMI-C+ medium. On day three, a pulse of ³H-thymidine was added, and the cells were harvested on day 4 for lysis and scintillation counting. Phytohemagglutinin (PHA) was used as a mitogen for positive control wells. A stimulation index was calculated for each antigen dosage level by dividing the counts per minute (cpm) of ³H-thymidine incorporated in the antigen-stimulated cells by the cpm in a control (unstimulated) well.

Following are examples which illustrate procedures for practicing the invention. These examples should not be construed as limiting. All percentages are by weight and all solvent mixture proportions are by volume unless otherwise noted.

Example 1 – *In vitro* studies in murine C2C12 myoblasts

In order to determine the relative strength of a number of constitutively active promoters in the context of AAV-AAT vectors, packageable AAV-AAT expression vectors containing one of the CMV, EF1, Ula or Ulb promoters (Figure 1) were constructed. Each of these constructs were transfected in to the murine C2C12 myoblast cell line. Both the EF1 and the CMV promoter were active for AAT expression, with EF1 construct (AAV-E-AT) expressing 850 ng/10⁵ cells/day and the CMV construct (AAV-C-AT) expressing approximately 670 ng/10⁵ cells/day, as measured by a human-specific ELISA assay for AAT (Figure 2). This difference was not statistically significant. The levels of expression from the Ula and Ulb constructs were undetectable.

In order to better characterize the level and duration of expression in the setting of vector transduction, cultures of C2C12 cells were transduced with either AAV-E-AT

or AAV-C-AT at multiplicities of infection ranging from 4×10^5 to 4×10^6 Dnase-resistant particles per cell. Cells were then selected for expression of the *neo* gene (present in each of the AAV constructs) by growth in G418-containing medium. Several cell clones and pooled cell populations were independently analyzed for AAT expression at four weeks post-transduction (Figure 3). There was a clear trend toward higher levels of expression at higher multiplicities of infection, and the E-AT construct expressed at least 10-fold greater quantities under all conditions in these long-term cultures. The most active E-AT clone expressed hAAT at a rate of over 1400 ng/ 10^5 cells/day.

Example 2—*In vivo* expression of hAAT from murine skeletal muscle

In order to determine whether the AAV-AAT constructs would be active *in vivo* in skeletal muscle, doses of vector were injected into the quadriceps femoris muscle of mice. Circulating serum levels of hAAT were then measured for 11 to 15 weeks after the initial injection. Four saline-injected animals from each mouse strain served as controls. In the case of the C-AT vector (Figure 5A), levels of expression were sufficient to achieve serum levels in excess of 800 μ g/ml in SCID mice after a single injection of 1.4×10^{13} particles. A dose-effect relationship was observed, with expression levels in SCID being at least 20-fold lower at the 5×10^{11} particle dose. The levels of expression increased over the first several weeks after injection and were stable thereafter until the time of sacrifice. Since hAAT has a half-life of less than 1 week, this indicated continuous expression. Levels from C57B1/6 mice were comparable, and also achieved values close to the therapeutic range. In similar studies, two of three Balb/c mice injected with 1×10^{11} particles of the C-AT vector did not express hAAT at detectable levels. Both of these were found to have developed high levels of anti-hAAT antibodies.

Surprisingly, expression levels from the AAV-E-AT vector after *in vivo* injection were modestly lower than those seen with the C-AT vector (Figure 5B), with maximal levels of approximately 250 ng/ml at the 5×10^{11} dose at and beyond 7 weeks in SCID mice. When the dose was further increased to 1×10^{12} particles, levels of approximately 1200 ng/ml were observed. These levels were stable for one year post-injection (Figure 5C). Levels observed in SCID and immune competent C57B1/6 mice were similar.

Example 3 — Immunologic Studies

In studies in Balb/c mice, antibody levels against hAAT were high in 2 of 3 animals injected. The one which did not have circulating anti-hAAT was the only animal with levels of hAAT expression similar to those in the C57B1/6 and SCID groups. The high-dose C57-C-AT injection group had detectable levels of antibody directed against VP3, but not hAAT.

In order to determine whether any cell-mediated immune responses were mounted, lymphocyte proliferation assays were performed using either hAAT or AAV-VP3 for antigenic stimulation of primary splenic lymphocytes harvested at the time of animal sacrifice, 16 weeks post-vector injection. Using this method, no immune responses were detectable in any of the mice.

Example 4 — Lack of toxicity from direct vector injection

In order to determine whether there was any direct toxicity, inflammation, or neoplastic change associated with vector injection, animals underwent complete necropsies. Histopathologic examination was performed on 5 μ m sections taken from the site of vector injection and from a panel of other organs, including the brain, heart, lungs, trachea, pancreas, spleen, liver, kidney, and jejunum. No histologic abnormalities were observed in any of these sites, even among those mice which developed humanol immune responses against hAAT.

Example 5 — Molecular evidence of AAV-AAT vector persistence

To confirm the presence of vector DNA, a vector-specific PCR (*neo* primers 5'-TATGGGATCGGCCATTGAAC-3', and 5'-CCTGATGCTCTTC-GTCCAGA-3', was performed on DNA extracted from 3 SCID mice 16 weeks after injection with the C-AT vector, and PCR products were analyzed by Southern blot analysis with a ³²P-labeled vector-specific probe (Figure 8). The state of vector DNA was analyzed using the Hirt procedure (Carter *et al*, 1983) to separate the low molecular weight episomal DNA from the high molecular weight fraction, which would contain integrated forms and large concatemers. In each case, vector DNA was present in the high molecular weight DNA fraction, whereas in only one of the animals was there a signal in the episomal fraction. This result indicates that by 16 weeks most of the vector DNA in our animals was either integrated or in large concatemers.

Example 6 — *In vivo* expression of hAAT from murine liver

Portal vein or tail vein injections were performed on 18 female C57BL/6 mice 8-10 weeks of age. The injection volume was 100 μ l per mouse.

Each group had the following parameters:

- 5 1. Group 1: 100 μ l of PBS n=4.
2. Group 2: 100 μ l of p43CB-AT (3×10^{10} IU/animal) n=3.
3. Group 3: 100 μ l of p43CB-AT (4×10^9 IU/animal) n=4.
4. Group 4: 100 μ l of C-AT (4×10^9 IU/animal) n=2.
5. Group 5: 100 μ l of E-AT (4×10^9 IU/animal) n=4.
- 10 6. Group 6: EATM TV=100 μ l by tail vein injection of E-AT (4×10^9 IU/animal) n=3.
7. Group 0: 100 μ l of PBS by tail vein injection n=2.

A total of 22 animals were used in this study.

15 All animals were anesthetized with 2-2-2 tribromoethanol (Avertin) using a working solution of 20 mg/ml at a dosage of 0.5 mg/g IP. A 2 cm ventral midline abdominal incision was made from the pubic symphysis extending cranially to the xyphoid process through skin and muscle layers. The portal vein was exposed by retracting the intestines and associated mesentery to the left side of the animal. Additionally, the quadrate and right medial lobes of the liver were retracted cranially.

20 Intestines and peritoneal cavity were continuously lavaged with 0.9% NaCl.

 Virus or PBS was delivered into the portal vein using a 30 g needle attached to a 100 μ l capillary pipette using mouth delivery via rubber tubing and a Drummond self-locking double layer 0.8 μ m filter. A small piece of Gel-Foam (.5x.5cm) was applied to the injection site before the needle was removed from the portal vein. The needle was

25 retracted from beneath the Gel-Foam and the piece was held in place with forceps while the intestines were replaced into the peritoneal cavity.

 The muscle and skin were closed in one layer using 2 simple interrupted 3-0 nylon sutures on an FS-1 cutting needle. Surgeries were performed on a thermoregulated operating board designed to maintain a temperature of 37 degrees. For recovery from

30 anesthesia, the animals were placed under a heat lamp adjusted to maintain an ambient temperature of approximately 37 degrees and given subcutaneous fluid if there was a significant amount of blood loss during surgery.

Serum levels of hAAT in the mice were measured two weeks after injection. Serum levels of about 200-150 $\mu\text{g/ml}$ hAAT were detected in mice receiving the p43CB-AT vector (Figure 13). Studies using the E-AT vector show that injection of vector by portal vein led to greater levels of hAAT secretion as compared to E-AT administered by tail vein injection.

Example 7—*In vivo* expression of hAAT from murine lung

Mice were injected intratracheally with either C-AT or p43CB-AT vector. Serum levels of hAAT in the mice were measured at day 3, 14 and 31 after injection (Figure 14). The p43CB-AT vector mediated high levels of expression of hAAT in lung.

It should be understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and the scope of the appended claims.

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We claim:

Claims

1 1. A method for providing an animal with a therapeutically effective amount of
2 a serum protein, said method comprising introducing into cells of said animal an effective
3 amount of viral particles or vector, wherein said viral particles or viral vector comprises
4 a polynucleotide encoding said protein.

1 2. The method according to claim 1, wherein said animal is a mammal.

1 3. The method according to claim 2, wherein said mammal is a human.

1 4. The method according to claim 1, wherein said vector is an adeno-associated
2 virus vector.

1 5. The method according to claim 1, wherein said vector comprises a promoter
2 sequence capable of driving expression of said polynucleotide encoding said protein.

1 6. The method according to claim 5, wherein said promoter sequence is selected
2 from the group consisting of CMV promoter sequences, hybrid CMV enhancer/ β -actin
3 promoter sequences, EF1 promoter sequences, Ula promoter sequences and U1b
4 promoter sequences.

1 7. The method according to claim 5, wherein said promoter sequence is an
2 inducible promoter selected from the group consisting of Tet-inducible promoters and
3 VP16-LexA promoters.

1 8. The method according to claim 5, wherein said vector further comprises an
2 enhancer sequence.

1 9. The method according to claim 8, wherein said enhancer is a synthetic
2 enhancer.

1 10. The method according to claim 1, wherein said animal has a condition that
2 results in a defective protein or a deficiency of said protein encoded by said
3 polynucleotide.

1 11. The method according to claim 1, wherein said animal has a condition that
2 can be ameliorated or treated by said protein encoded by said polynucleotide.

1 12. The method according to claim 1, wherein said protein encoded by said
2 polynucleotide is selected from the group consisting of anti-proteases, enzymes,
3 structural proteins, coagulase factors, interleukins, cytokines, growth factors, interferons,
4 and lymphokines.

1 13. The method according to claim 1, wherein said cells are myofibers,
2 myoblasts, hepatocytes, or lung cells.

1 14. The method according to claim 1, wherein said polynucleotide encodes
2 human alpha-1-antitrypsin protein, or a biologically active fragment or variant thereof.

1 15. The method according to claim 4, wherein said polynucleotide encodes
2 human alpha-1-antitrypsin protein, or a biologically active fragment or variant thereof.

1 16. The method according to claim 1, wherein said viral particles are introduced
2 into said cells or tissue by infection or injection.

1 17. The method according to claim 1, wherein said vector is introduced into said
2 cells by transfection or injection.

1 18. The method according to claim 1, wherein said viral particles or vector is
2 introduced into said cells *in vitro* and said treated cells are introduced into said animal.

1 19. The method according to claim 1, wherein said viral particles or vector is
2 introduced into said cells *in vivo*.

1 20. The method according to claim 19, wherein said viral particles or vector is
2 injected into muscle.

1 21. The method according to claim 19, wherein said viral particles or vector is
2 injected into portal or peripheral vein.

1 22. The method according to claim 19, wherein said viral particles or vector is
2 injected intratracheally or inhaled into the lungs.

1 23. The method according to claim 15, wherein said vector is selected from the
2 group consisting of dE-AT, E-AT, C-AT, C-AT2, p43C-AT, p43CB-AT, p43C-AT-IN,
3 p43msENC-AT, p43rmsENC-AT, p43msENCB-AT and p43rmsENCB-AT.

1 24. A recombinant viral vector comprising a polynucleotide encoding a protein
2 capable of providing a therapeutic effect to an animal when expressed in said animal.

1 25. The vector according to claim 24, wherein said animal is a mammal.

1 26. The vector according to claim 25, wherein said mammal is a human.

1 27. The vector according to claim 26, wherein said vector is an adeno-associated
2 virus vector.

1 28. The vector according to claim 24, wherein said vector comprises a promoter
2 sequence capable of driving expression of said polynucleotide encoding said protein.

1 29. The vector according to claim 24, wherein said promoter sequence is selected
2 from the group consisting of CMV promoter sequences, hybrid CMV enhancer/ β -actin

3 promoter sequences, EF1 promoter sequences, Ula promoter sequences and U1b
4 promoter sequences.

1 30. The vector according to claim 24, wherein said polynucleotide encodes
2 human alpha-1-antitrypsin protein, of a biologically active fragment or variant thereof.

1 31. The vector according to claim 27, wherein said polynucleotide encodes
2 human alpha-1-antitrypsin protein, of a biologically active fragment or variant thereof.

1 32. The vector according to claim 31, wherein said vector is selected from the
2 group consisting of dE-AT, E-AT, C-AT, C-AT2, p43C-AT, p43CB-AT, p43C-AT-IN,
3 p43msENC-AT, p43rmsENC-AT, p43msENCB-AT and p43rmsENCB-AT.

1 33. A viral particle comprising the vector of claim 24.

1 34. A cell comprising the vector of claim 24.

1 35. The cell according to claim 34, wherein said cell is a myofiber, myoblast,
2 hepatocyte, or lung cell.

1 36. A method for treating alpha-1-antitrypsin deficiency in an animal, said
2 method comprising introducing into cells of said animal a vector according to claim 24,
3 wherein said polynucleotide of said vector encodes alpha-1-antitrypsin protein, or a
4 biologically active fragment or variant thereof.

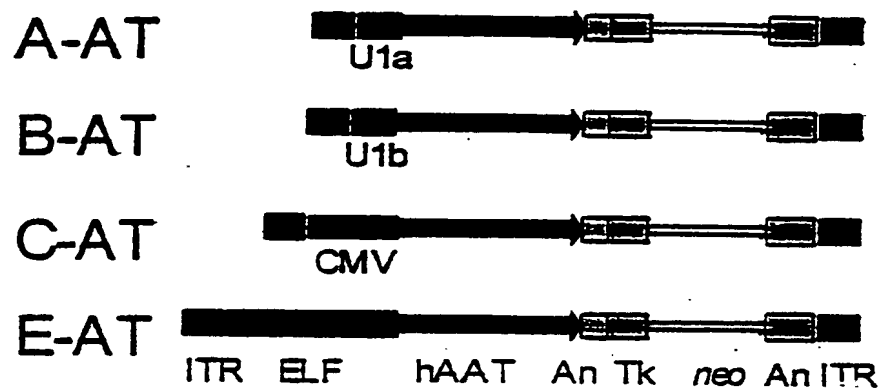


FIGURE 1

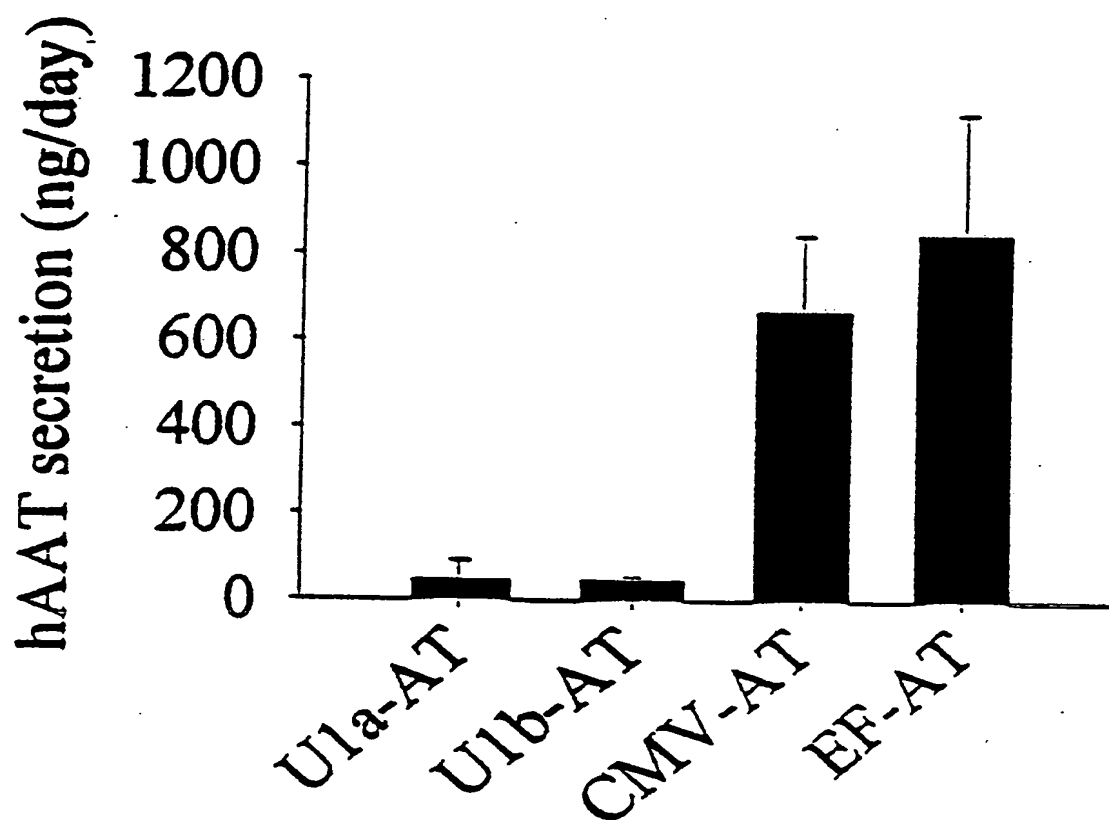


FIGURE 2

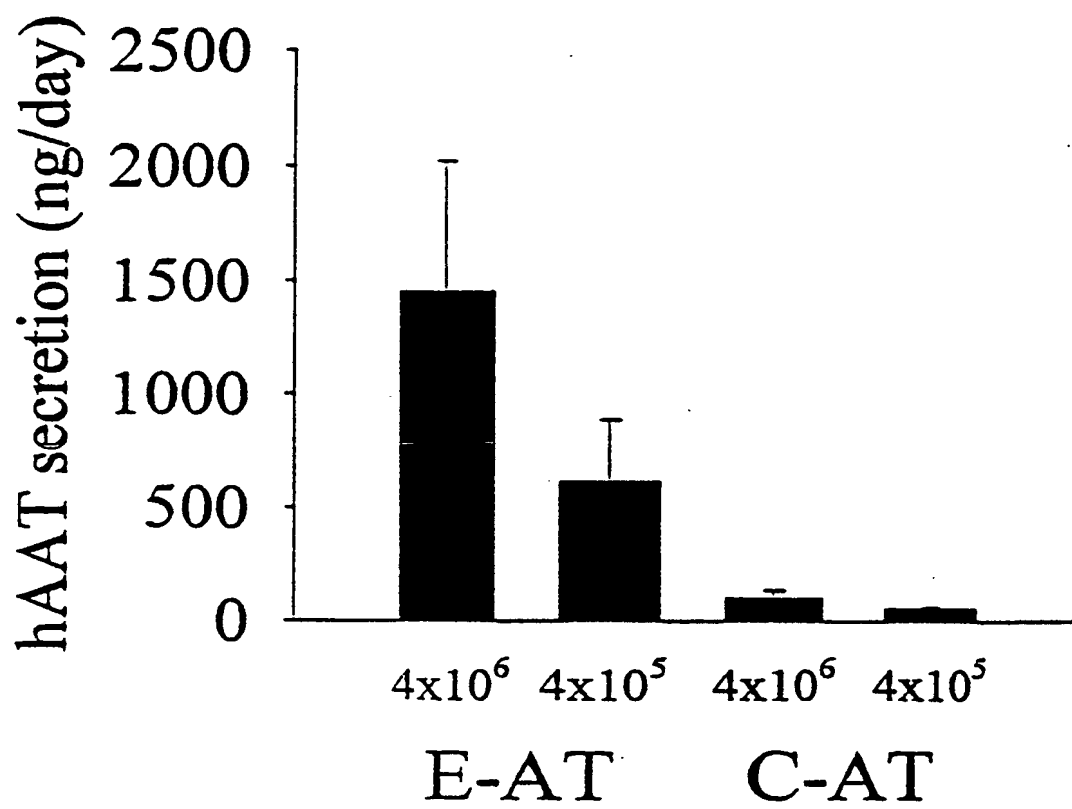


FIGURE 3

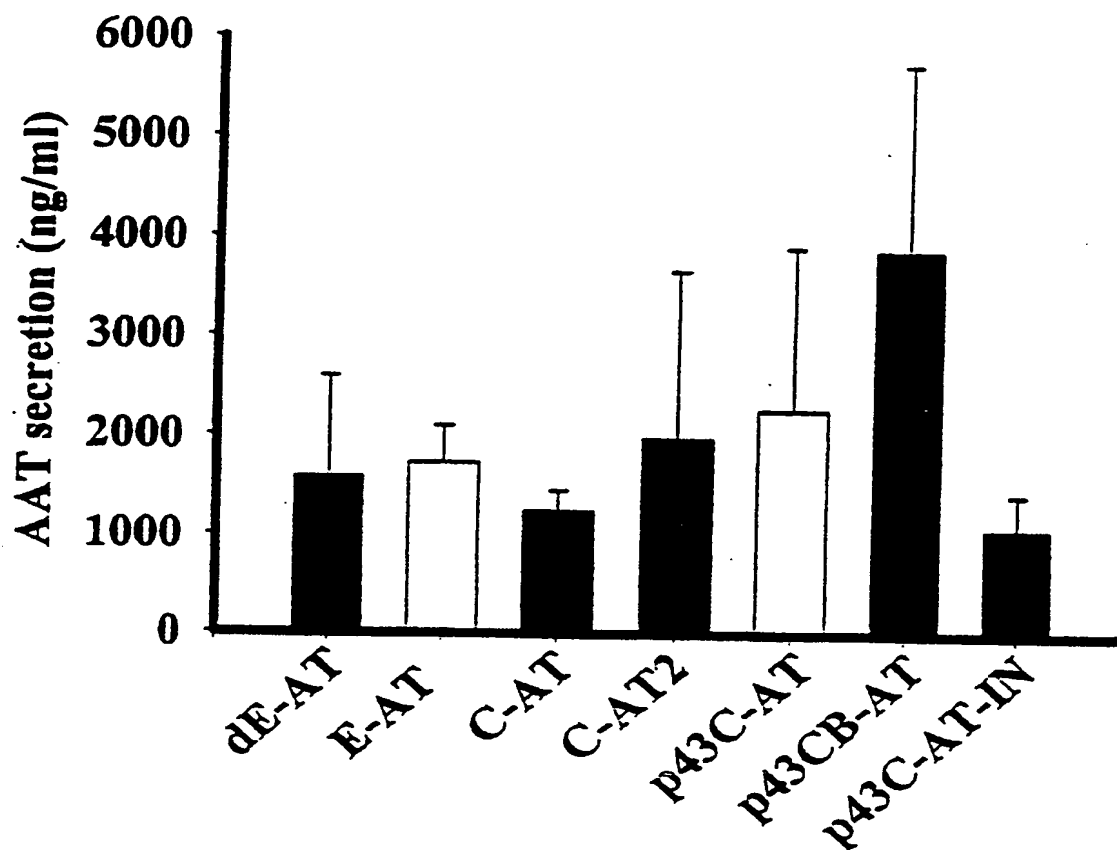


FIGURE 4

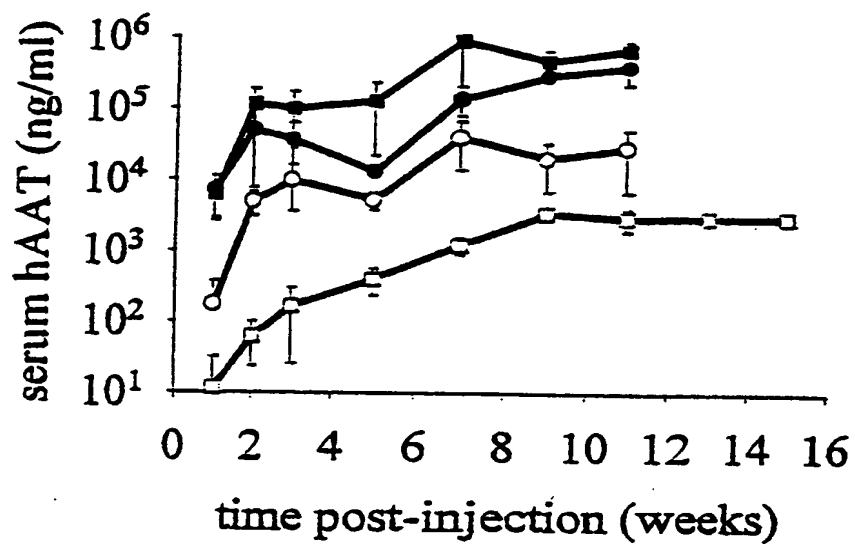


FIGURE 5A

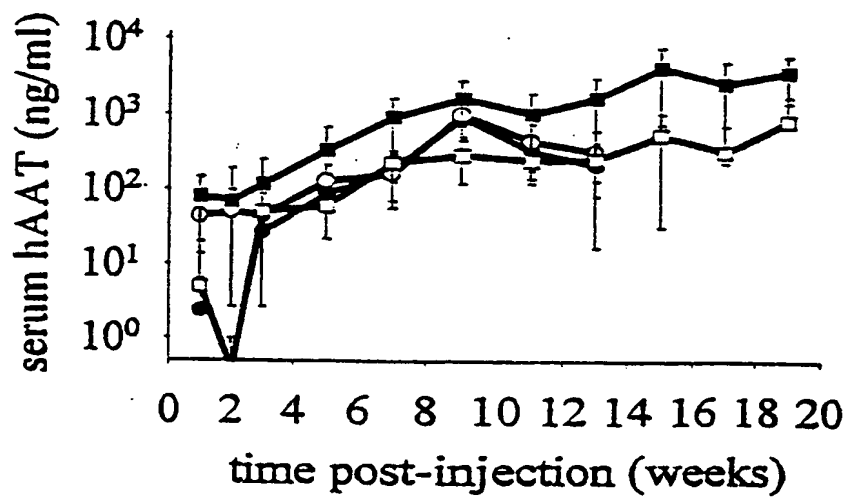


FIGURE 5B

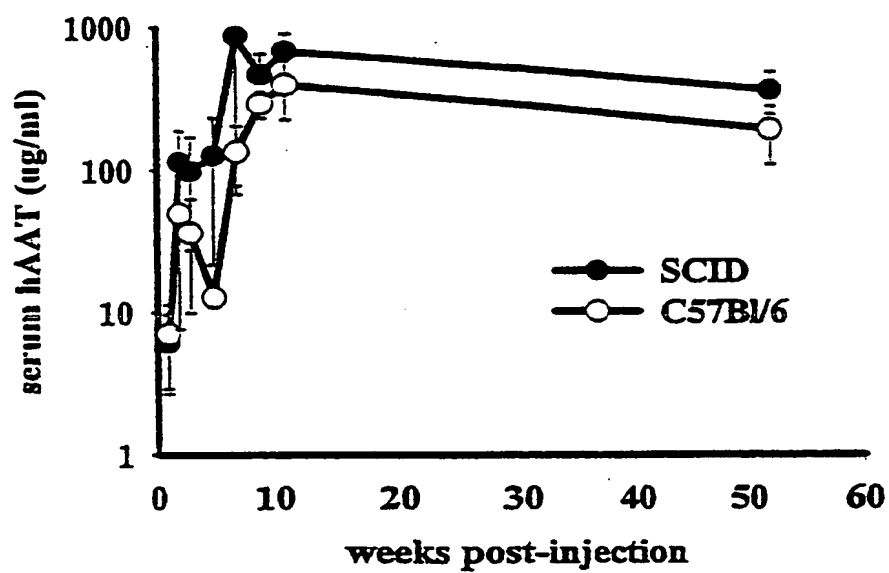


FIGURE 5C

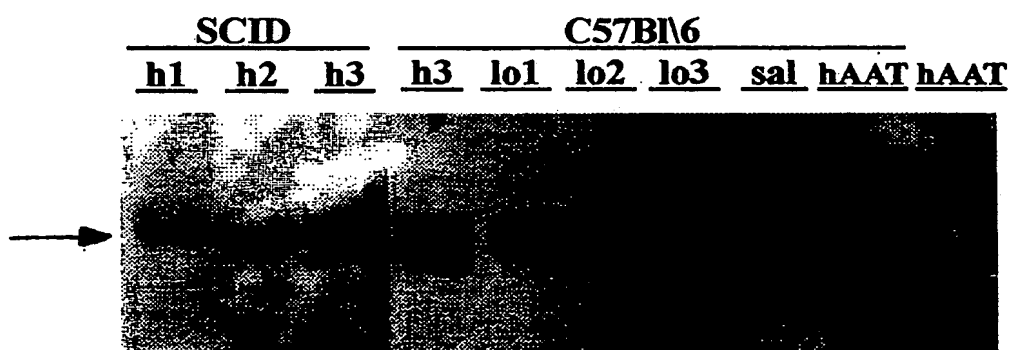


FIGURE 6

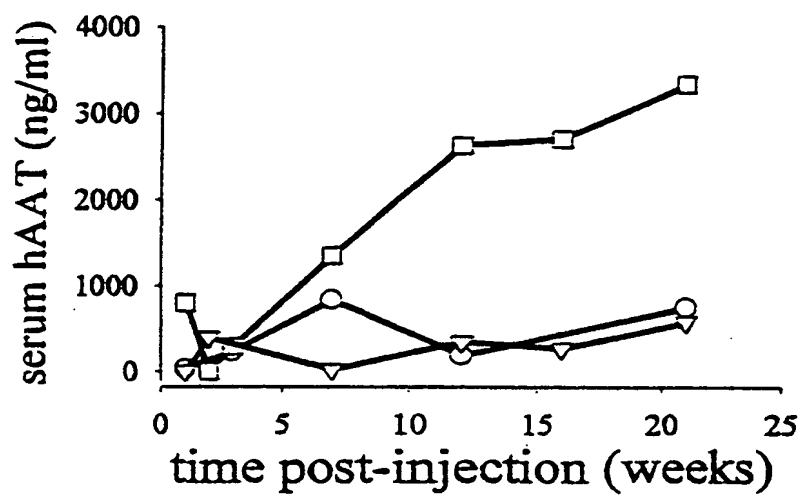


FIGURE 7A

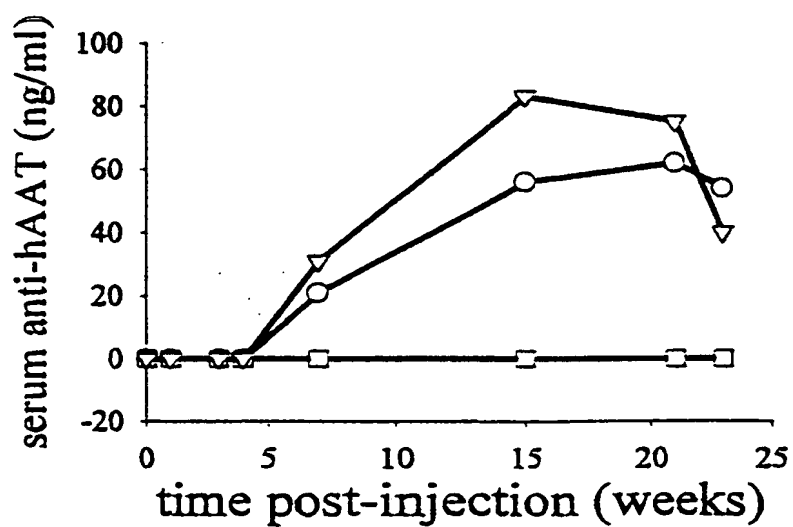


FIGURE 7B

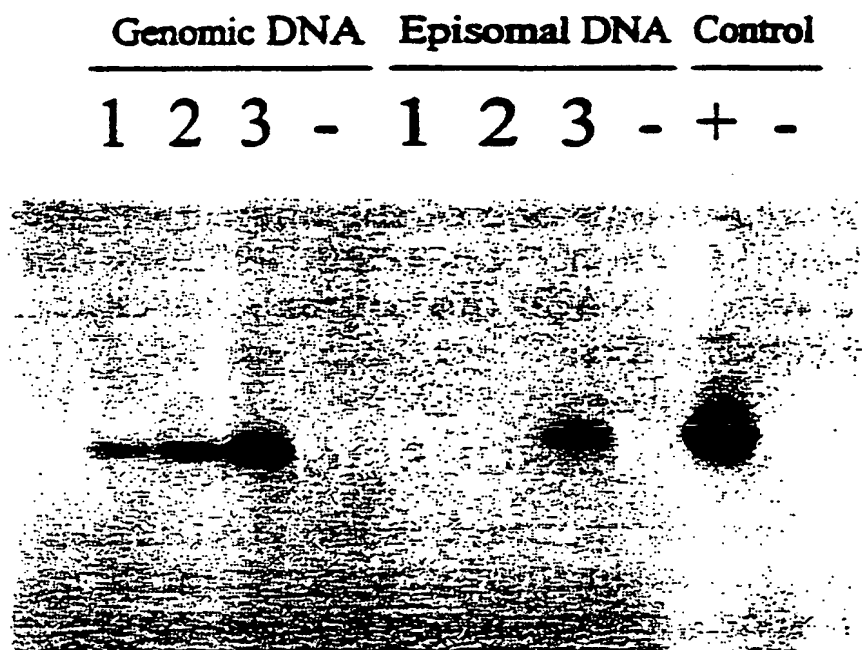


FIGURE 8

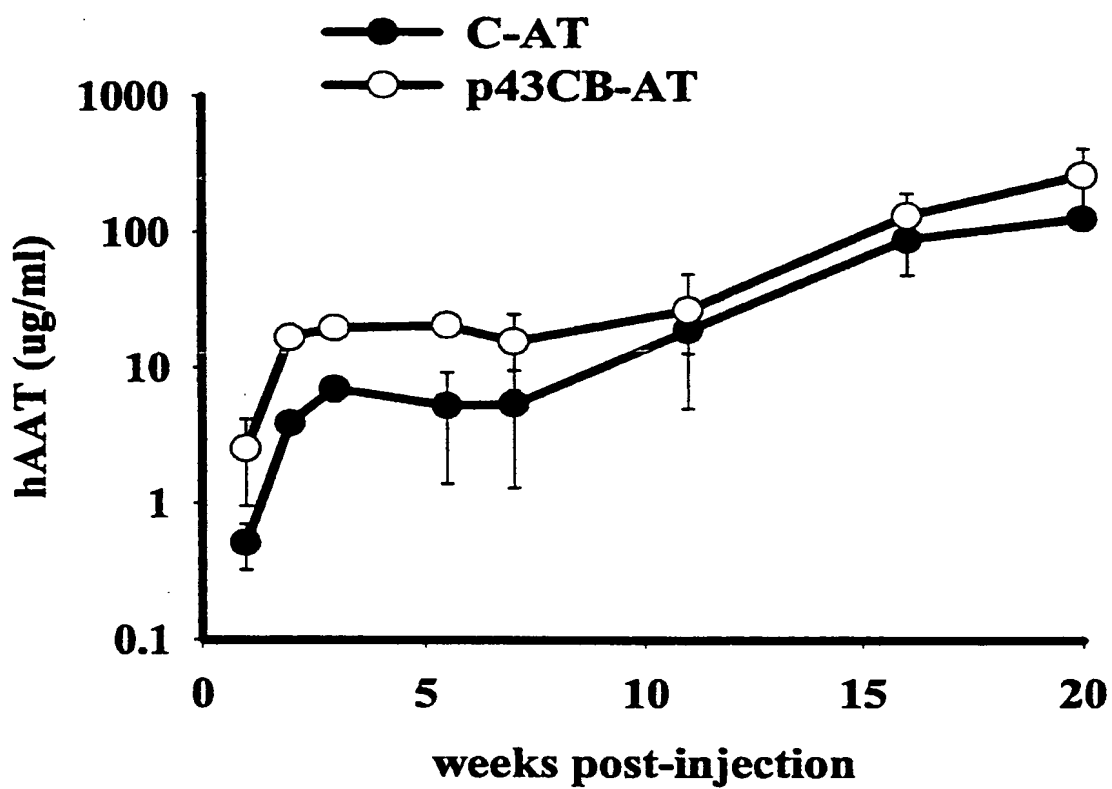


FIGURE 9

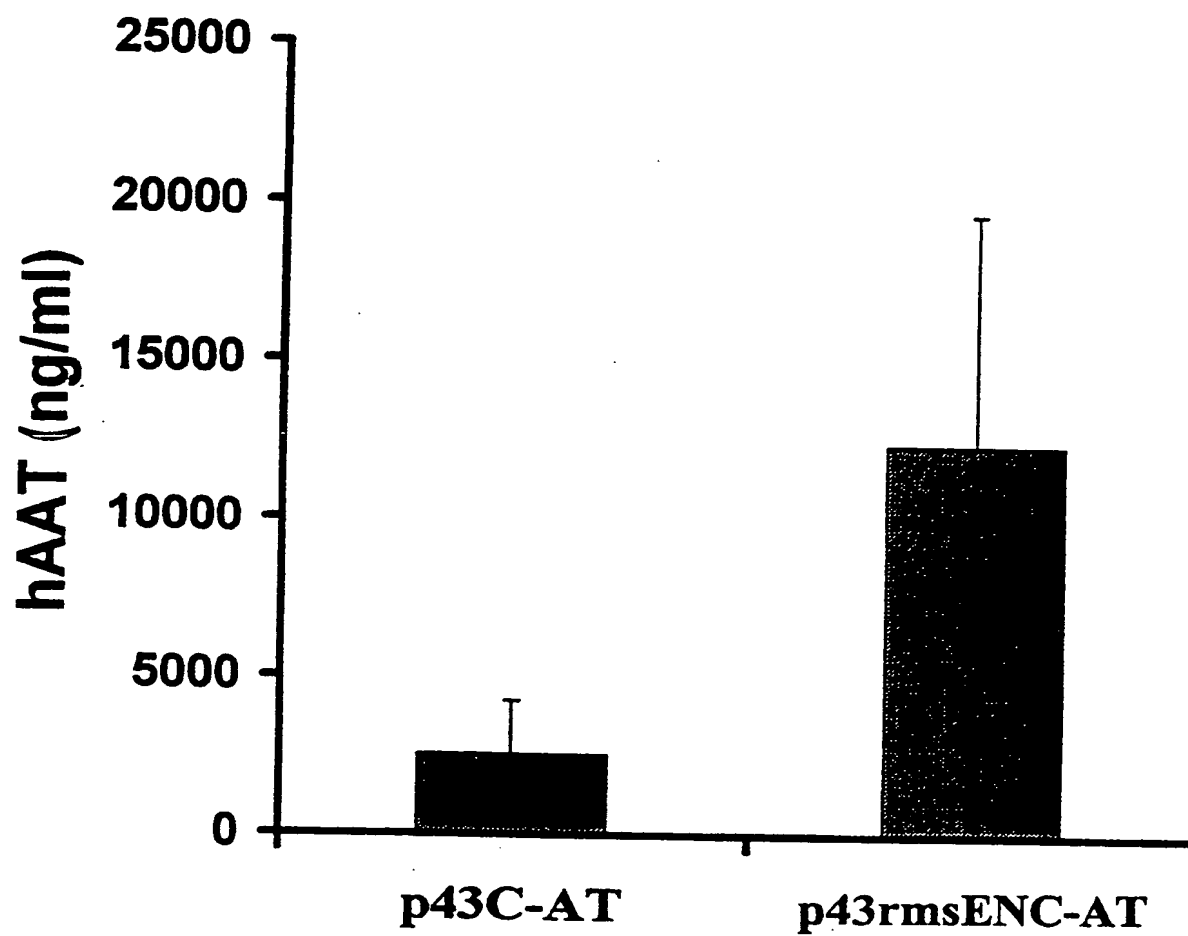


FIGURE 10

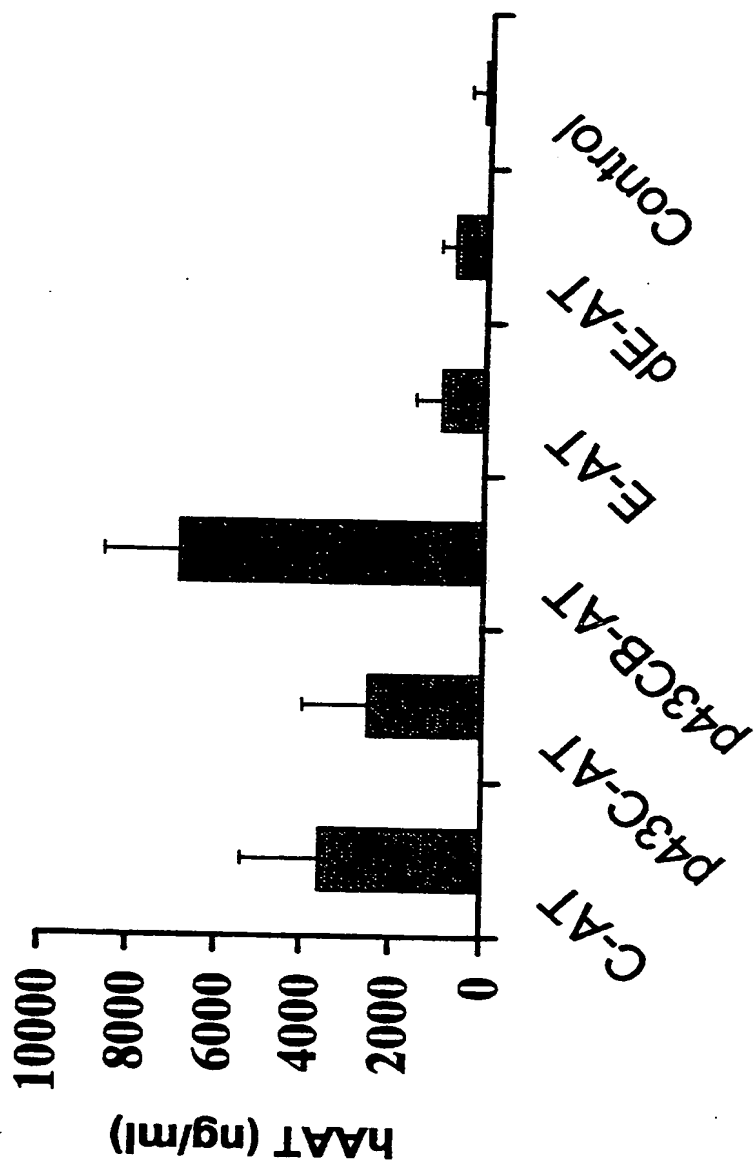


FIGURE 11

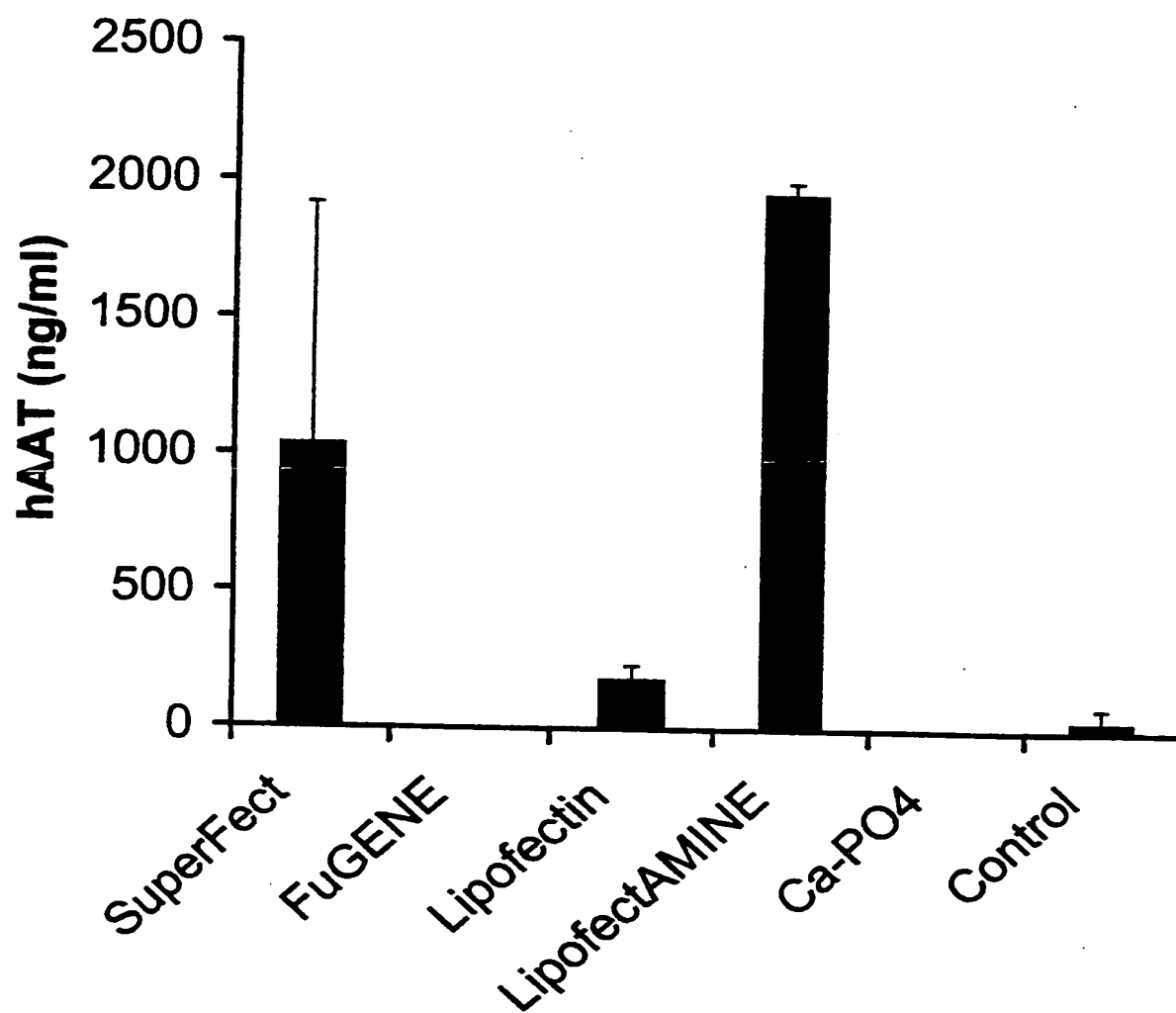


FIGURE 12

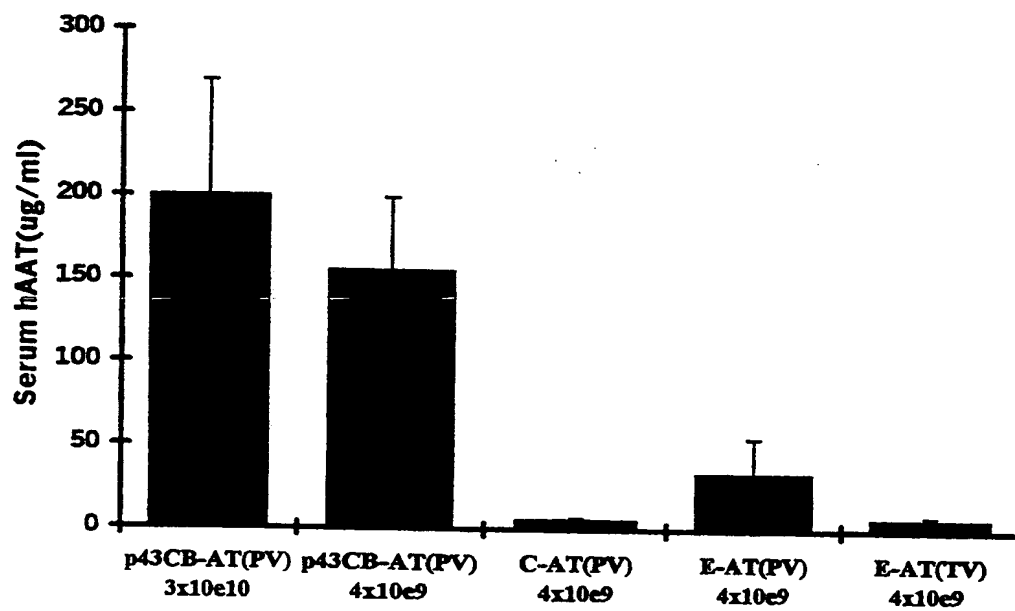


FIGURE 13

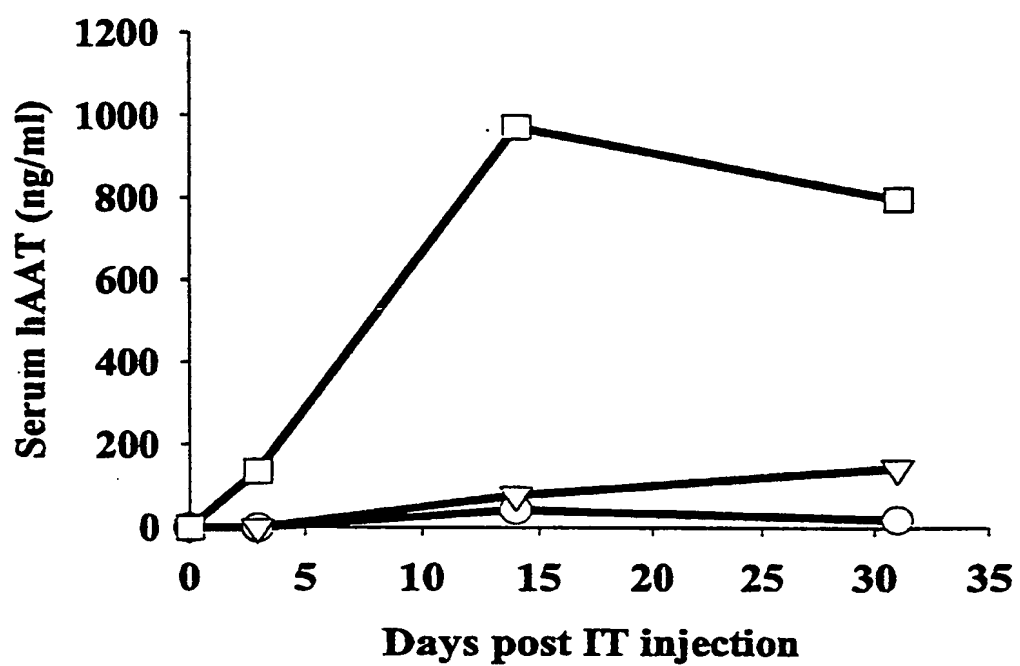


FIGURE 14

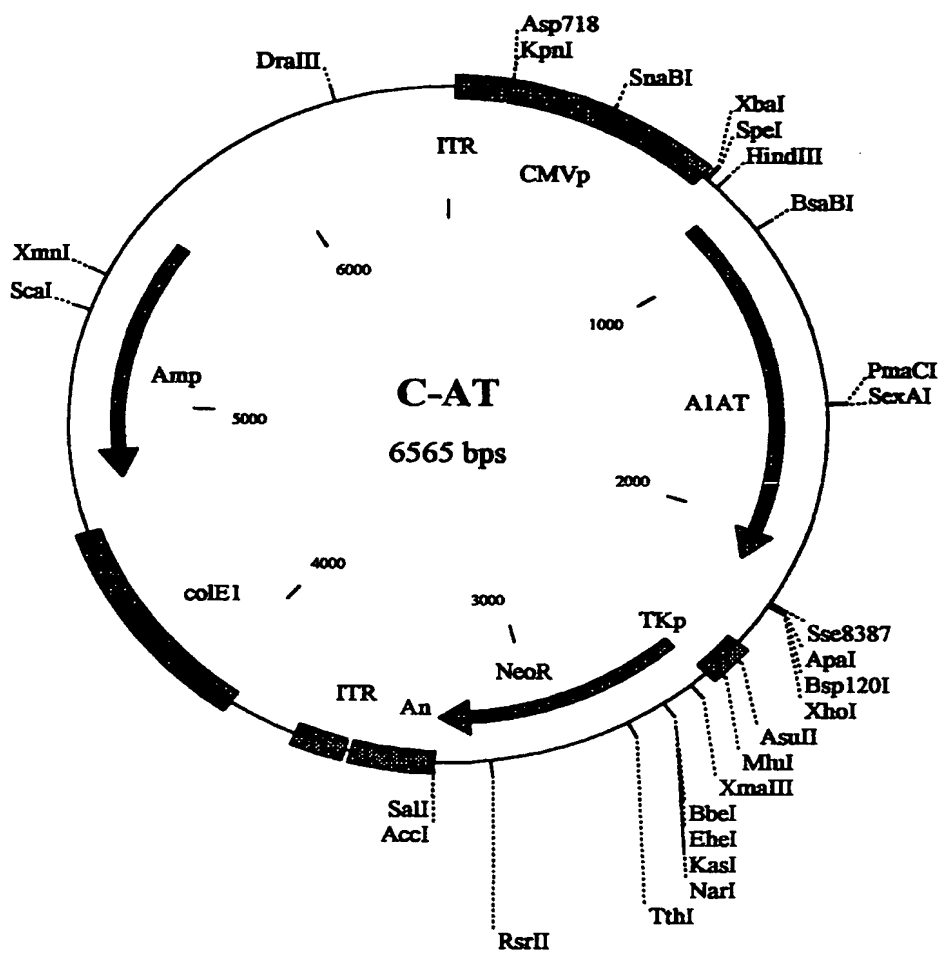


FIGURE 15

Molecule Name: C-AT 6565 bps DNA Circular
Sequence Printed: 1-6565 (Full) Date Printed 16 Apr 1999
Description: Ligation of pTR and aat

```
1  gggggggggg gggggggggtt ggccactccc tctctgcgcg ctcgctcgct
51  cactgaggcc gggcgaccaa aggtcgcccg acgcccgggc tttgcccggg
101 cggcctcagt gagcgagcga gcgcgcagag agggagtggc caactccatc
151 actaggggtt cctagatctg aattcggtac ccgttacata acttacggta
201 aatggccccg ctggctgacc gcccaacgac ccccgcccat tgacgtcaat
251 aatgacgtat gttcccatag taacgccaat agggactttc cattgacgtc
301 aatgggtgga gtatttacgg taaactgccc acttggcagt acatcaagtg
351 tatcatatgc caagtacgcc ccctattgac gtcaatgacg gtaaattggc
401 cgcctggcat tatgcccagt acatgacctt atgggacttt cctacttggc
451 agtacatcta cgtattagtc atcgctatta ccatgggtgat gcggttttgg
501 cagtacatca atgggctggg atagcggttt gactcacggg gatttccaag
551 tctccacccc attgacgtca atgggagttt gttttggcac caaatcaac
601 gggactttcc aaaatgtcgt aacaactccg cccattgac gcaaattggc
651 ggtaggcgtg tacggtggga ggtctatata agcagagctc gtttagtgaa
701 ccgtcagatc gcctggagac gccatccacg ctgttttgac ctccatagaa
751 gacaccggga ccgatccagc ctccggactc tagaactagt ggatcccccg
801 ggctgcagga attcgatatc aagcttgggg attttcaggc accaccactg
851 acctgggaca gtgaatcgac aatgccgtct tctgtctcgt ggggcatcct
901 cctgctggca ggctgtgct gcctgggtcc tgtctccctg gctgaggatc
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1251 tcaaccagcc agacagccag ctccagctga ccaccggcaa tggcctgttc
1301 ctcagcgagg gcctgaagct agtgggataag tttttggagg atgttaaaaa
1351 ttgttaccac tcagaagcct tcactgtcaa cttcggggac accgaagagg
1401 ccaagaaaca gtcaacgat tacytgaga agggtaactc agggaaattt
1451 gtggatttgg tcaaggagct tgacagagac acagtttttg ctctggtgaa
1501 ttacatcttc tttaaaggca aatgggagag accctttgaa gtcaaggaca
1551 ccgaggaaga ggacttcac gtggaccagg tgaccaccgt gaagggtgct
1601 atgatgaagc gtttaggcat gtttaacatc cagcactgta agaagctgtc
1651 cagctgggtg ctgctgatga aatacctggg caatgccacc gccatcttct
1701 tctgcctga tgaggggaaa ctacagcacc tggaaaatga actcaccac
1751 gatcatcatc ccaagtctct ggaaaatgaa gacagaaggt ctgccagctt
1801 acattttacc aaactgtcca ttactggaac ctatgatctg aagagcgtcc
1851 tgggtcaact gggcatcact aaggtcttca gcaatggggc tgacctctcc
1901 ggggtcacag aggaggcacc cctgaagctc tccaaggccg tgcataaggc
1951 tgtgtgacc atcgacgaga aagggactga agctgctggg gccatgtttt
2001 tagaggccat acccatgtct atcccccccg aggtcaagtt caacaaaacc
2051 tttgtcttct taatgattga acaaaaatac aagtctcccc tcttcatggg
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2201 ctggtaaccc ccccccccc tgcaagggcc ctcgagcagt gtggttttgc
2251 aagaggaagc aaaaagcctc tccaccagg cctggaatgt ttccacccaa
2301 gtcgaaggca gtgtgggttt gcaagaggaa gcaaaaagcc tctccaccca
2351 ggcttggaa gtttccaccc aatgtcgagc aaccccgccc agcgtcttgt
2401 cattggcgaa ttcgaacacg cagatgcagt cggggcgggc cgggtcccagg
2451 tccacttcgc atattaaggt gacgcgtgtg gcctcgaaca ccgagcgacc
2501 ctgcagccaa tatgggatcg gccattgaac aagatggatt gcacgcaggt
2551 tctccggccg cttgggtgga gaggtatttc ggctatgact gggcacaaca
2601 gacaatcggc tgctctgatg ccgccgtgtt ccggtgttca gcgcaggggc
2651 gcccggttct ttttgtcaag accgacctgt ccggtgccct gaatgaactg
2701 caggacgagg cagcgcggct atcgtggctg gccacgacg gcgttccttg
```

FIGURE 15A

```

2751 cgcagctgtg ctccgacgttg tcaactgaagc ggggaagggac tggctgctat
2801 tgggcggaagt gccgggggcag gatctcctgt catctcacct tgctcctgcc
2851 gagaaagtat ccatcatggc tgatgcaatg cggcggtctgc atacgcttga
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2951 cacgtactcg gatggaagcc ggtcttgtcg atcaggatga tctggacgaa
3001 gagcatcagg ggctcgcgcc agccgaactg ttcgccaggc tcaaggcgcg
3051 catgccccgac ggcgaggatc tcgtcgtgac ccatggcgat gcctgcttgc
3101 cgaatatcat ggtggaaaat ggccgctttt ctggattcat cgactgtggc
3151 cggctgggtg tggcggaccg ctatcaggac atagcggttg ctaccctgta
3201 tattgctgaa gagcttggcg gcgaatgggc tgaccgcttc ctctgtcttt
3251 acggtatcgc cgctcccgat tcgcagcgca tcgccttcta tcgccttctt
3301 gacgagttct tctgagggga tccgtcgact agagctcgct gatcagctc
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3651 cgggcgacct ttggtcgccc ggcctcagtg agcgagcgag cgcgacagga
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3751 tcggccaacg cgcggggaga ggcgggttgc gtattgggcg ctcttccgct
3801 tcctcgctca ctgactcgct gcgctcggtc gttcggctgc ggcgagcggt
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3901 acgcaggaaa gaacatgtga gcaaaaggcc agcaaaaggc caggaccagt
3951 aaaaaggccg cgttgcctggc gtttttccat aggtcccgcc cccctgacga
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4051 tataaagata ccaggcggtt cccctggaa gctccctcgt gcgctctcct
4101 gttccgaccc tgccgcttac cggataacctg tccgcctttc tccctcggg
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4201 aggtcgctcg ctccaagctg ggctgtgtgc acgaaccccc cgttcagccc
4251 gactcccgcg ccttatccgg taactatcgt cttagtcca acccggttag
4301 acacgactta tcgccactgg cagcagccac tggtaacagg attagcagag
4351 cgaggatatg aggcgggtgt acagagttct tgaagtgggt gcctaactac
4401 ggctacacta gaaggacagt atttgggtatc tgcgctctgc tgaagccagt
4451 taccttcgga aaaagagttg gtagctcttg atccggcaaa caaaccaccg
4501 ctggtagcgg tgggtttttt gtttgcaagc agcagattac gcgcagaaaa
4551 aaaggatctc aagaagatcc tttgatcttt tctacggggt ctgacgctca
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4651 ggatcttcac ctagatcctt ttaaattaaa aatgaagttt taaatcaatc
4701 taaagtatat atgagtaaac ttgggtctgac agttaccaat gcttaatcag
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4851 cccagtgtgt caatgatacc gcgagacca cgctcaccgg ctccagattt
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5001 gtaagtagtt cgccagttaa tagtttgccg aacggtgttg ccattgctac
5051 aggcacgtg gtgtcacgct cgtcgttttg tatggcttca ttcagctccg
5101 gttcccaacg atcaaggcga gttacatgat ccccatggtt gtgcaaaaaa
5151 gcggttagct ccttcgggtcc tcgcatcggt gtcagaagta agttggccgc
5201 agtggttatc ctcatggtta tggcagcact gcataattct cttactgtca
5251 tgccatccgt aagatgcttt tctgtgactg gtgagtactc aaccaagtca
5301 ttctgagaat agtgtatgcg gcgaccgagt tgctcttgcc cggcgtaaat
5351 acgggataat accgcgccac atagcagaac tttaaaagtg ctcatcattg
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5501 tactttcacc agcgtttctg ggtgagcaaa aacaggaagg caaaatgccg
5551 caaaaaaggg aataaggcg acacggaaat gttgaatact catactcttc
5601 ctttttcaat attattgaag catttatcag ggttattgtc tcatgagcgg
5651 atacatattt gaatgtattt agaaaaataa acaaataggg gttccgcgca
5701 catttccccg aaaagtgcc cctgacgtct aagaaacat tattatcatg

```

FIGURE 15B

5751	acattaacct	ataaaaaatag	gcgtatcacg	aggccctttc	gtctcgcgcg
5801	tttcggtgat	gacggtgaaa	acctctgaca	catgcagctc	ccggagacgg
5851	tcacagcttg	tctgtaagcg	gatgccggga	gcagacaagc	ccgtcagggc
5901	gcgtcagcgg	gtgttggcgg	gtgtcggggc	tggcttaact	atgcggcatc
5951	agagcagatt	gtactgagag	tgcaccatat	gcggtgtgaa	ataccgcaca
6001	gatgcgtaag	gagaaaatac	cgcatacagga	aattgtaaac	gttaatatatt
6051	tgttaaaatt	cgcgttaaat	ttttgttaaa	tcagctcatt	ttttaaccaa
6101	taggccgaaa	tcggcaaaaat	cccttataaa	tcaaaagaat	agaccgagat
6151	agggttgagt	gttggtccag	tttggaaaca	gagtccacta	ttaaagaacg
6201	tggactccaa	cgtcaaaggg	cgaaaaaccg	tctatcaggg	cgatggccca
6251	ctacgtgaac	catcacccta	atcaagtttt	ttggggtcga	ggtgccgtaa
6301	agcactaaat	cggaacccta	aaggaggccc	ccgatttaga	gcttgacggg
6351	gaaagccggc	gaacgtggcg	agaaaggaag	ggaagaaagc	gaaaggagcg
6401	ggcgctaggg	cgctggcaag	tgtagcggtc	acgctgcgcg	taaccaccac
6451	acccgcccg	cttaatgcgc	cgctacaggg	cgcgtcgcgc	cattcgccat
6501	tcaggctacg	caactgttgg	gaagggcgat	cgggtgcgggc	ctcttcgcta
6551	ttacgccagg	ctgca			

FIGURE 15C

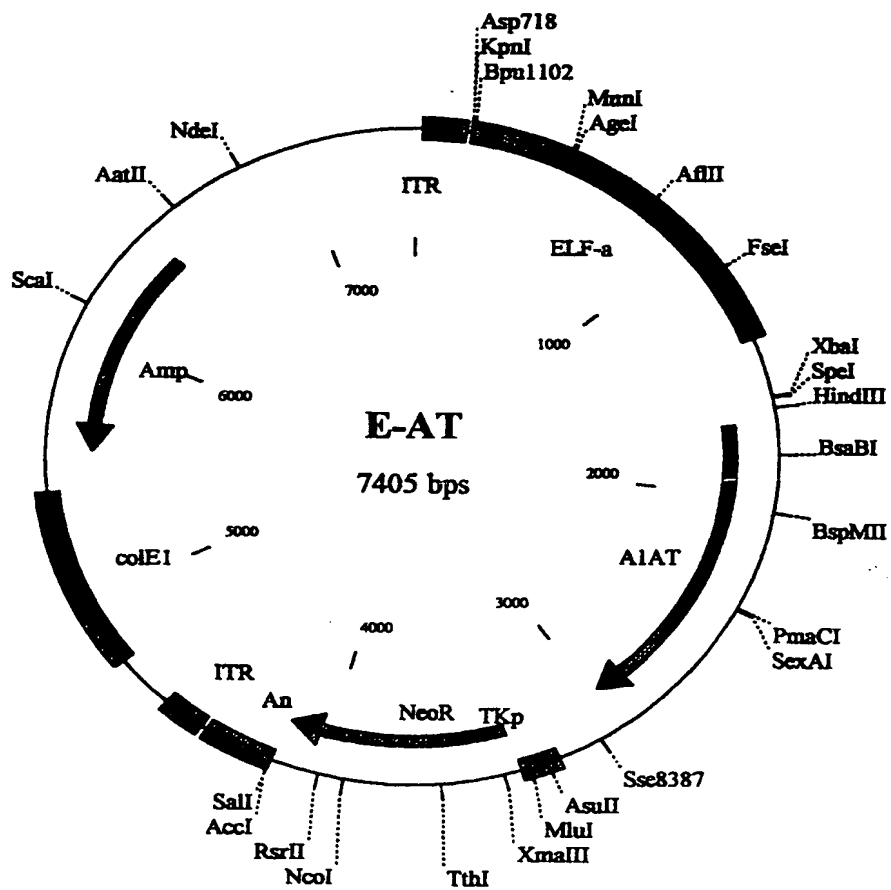


FIGURE 16

Molecule Name: E-A⁺ 7405 bps DNA Circular
Sequence Printed: 1-7405 (Full) Date Printed 16 Apr 1999
Description: Ligation of AAT and elf

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1  gggggggggg gggggggggt ggccactccc tctctgcgcg ctgcgtcgct
51  cactgaggcc gggcgaccaa aggtcgcccc acgcccgggc tttgcccggg
101 cggcctcagt gagcgagcga gcgcgcagag agggagtggc caactccatc
151 actaggggtt cctagatctg aattcggtac cttggagcta agccagcaat
201 ggtagaggga agattctgca cgtcccctcc aggcggcctc cccgtcacca
251 ccccccccaa cccgccccga ccggagctga gagtaattca tacaaaagga
301 ctgcgccctg ccttggggaa tcccagggaac cgtcggttaa ctcccactaa
351 cgtagaaccc agagatcgct gcgttccccg cccctcaccg gcccgtcttc
401 gtcatactg aggtggagaa gagcatgcgt gaggtccgg tgcccgtcag
451 tgggcagagc gcacatcgcc cacagtcccc gagaagttgg ggggaggggt
501 cggcaattga accggtgcct agagaaggtg gcgcggggta aactgggaaa
551 gtgatgtcgt gtactggctc cgcctttttc ccgagggtgg gggagaaccg
601 tatataagt cagtagtcgc cgtgaacgct ctttttcgca acgggtttgc
651 cgccagaaca caggtaatg ccgtgtgtgg ttcccgcggg cctggcctct
701 ttacgggtta tggcccttgc gtgccttgaa ttacttcac gccctggct
751 gcagtacgt attcttgat ccgagcttcg ggttggaggt ggggtggaga
801 gttcgaggcc ttgcgcttaa ggagcccctt cgcctcgtgc ttgagttgag
851 gcctggcctg ggcgtgggg ccgcgcgtg cgaatctggt ggcaccttcg
901 cgctgtctc gctgctttcg ataagtctct agccatttaa aatttttgat
951 gacctgctgc gacgcttttt ttctggcaag atagtcttgt aaatgcgggc
1001 caagatctgc acactggtat ttcggttttt ggggcgcgg gcggcgacgg
1051 ggcccgctgc tcccagcgca ctgttcggc gaggcggggc ctgagagcgg
1101 ggccaccgag aatcggaagg gggtagtctc aagctggccg gcctgctctg
1151 gtgcctggcc tcgcgcggcc gtgtatcgcc ccgcccggg cggcaaggct
1201 ggcccggtcg gcaccagttg cgtgagcgga aagatggccg cttcccggcc
1251 ctgctgcagc gagtcaaaa tggaggacgc ggcgctcggg agagcggcg
1301 ggtgagtcac ccacacaaag gaaaagggcc tttccgtcct cagccgtcgc
1351 ttcattgtac tccacggagt accgggcgcc gtccaggcac ctcgattagt
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1451 gcgatggagt ttccccacac ttgagtgggt gagactgaag ttaggccagc
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1551 cttggttcat tctcaagcct cagacagtgg ttcaaagttt ttttcttcca
1601 tttcaggtgt cgtgaaaatc tagaactagt ggatcccccg ggctgcagga
1651 attcgatatc aagcttgggg attttcaggc accaccactg acctgggaca
1701 gtgaatcgac aatgccgtct tctgtctcgt ggggcaccc cctgctggca
1751 ggctgtgct gccctggctcc tgtctccctg gctgaggatc cccagggaga
1801 tgctgcccag aagacagata catcccacca tgatcaggat caccacacct
1851 tcaacaagat ccccccaac ctggctgagt tcgccttcag cctataccgc
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2001 acgatgaaat cctggagggc ctgaatttca acctcacgga gattccggag
2051 gctcagatcc atgaaggctt ccaggaaact ctccgtacct tcaaccagcc
2101 agacagccag ctccagctga ccaccggcaa tggcctgttc ctcagcgagg
2151 gcctgaagct agtggataag tttttggagg atgttaaaaa gttgtaccac
2201 tcagaagcct tcaactgtca cttcggggac accgaagagg ccaagaaaca
2251 gatcaacgat tacgtggaga agggtaacta agggaaaatt gtggatttgg
2301 tcaaggagct tgacagagac acagtttttg ctctggtgaa ttacatcttc
2351 tttaaaggca aatgggagag accctttgaa gtcaaggaca ccgaggaaga
2401 ggacttccac gtggaccagg tgaccaccgt gaaggtgcct atgatgaagc
2451 gtttaggcat gtttaacatc cagcactgta agaagctgtc cagctgggtg
2501 ctgctgatga aatacctggg caatgccacc gccatcttct tcctgcctga
2551 tgaggggaaa ctacagcacc tggaaaatga actcaccac gatatcatca
2601 ccaagttcct ggaaaatgaa gacagaaggt ctgccagctt acatttacc
2651 aaactgtcca ttactggaac ctatgatctg aagagcgctc tgggtcaact
2701 gggcatcact aaggtcttca gcaatggggc tgacctctcc ggggtcacag
```

FIGURE 16A

```

2751 aggaggcacc cc aagctc tccaaggccg tgcataaggc tggctgacc
2801 atcgacgaga aagggaactga agctgctggg gccatgtttt tagaggccat
2851 acccatgtct atcccccccg aggtcaagtt caacaaaccc tttgtcttct
2901 taatgattga aaaaaataacc aagtctcccc tcttcatggg aaaagtgggtg
2951 aatcccaccc aaaaataact cctctcgct cctcaacccc tcccctccat
3001 ccctggcccc ctccctggat gacattaaag aagggttgag ctggtaaccc
3051 cccccccccc tgcaggggcc ctcgagcagt gtggttttgc aagaggaagc
3101 aaaaagcctc tccaccagc cctggaatgt ttccacccaa gtcgaaggca
3151 gtgtggtttt gcaagaggaa gcaaaaagcc tctccacca ggctggaat
3201 gtttccaccc aatgtcgagc aaccccgccc agcgtcttgt cattggcgaa
3251 ttcgaacacg cagatgcagt cggggcggcg cggctccagg tccacttcgc
3301 atattaaggt gacgcgtgtg gcctcgaaca ccgagcgacc ctgcagccaa
3351 tatgggatcg gccattgaac aagatggatt gcacgcaggt tctccggccg
3401 cttgggtgga gaggtattc ggctatgact gggcacaaca gacaatcggc
3451 tgctctgatg ccgctgtgtt ccggtgtca gcgcaggggc gcccggttct
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3601 ctcgacgttg tcaactgaagc gggaaggac tggctgctat tgggcgaagt
3651 gccggggcag gatctcctgt catctcacct tgctcctgcc gagaaagtat
3701 ccatcatggc tgatgcaatg cggcggctgc atacgcttga tccggtacc
3751 tgcccattcg accaccaagc aacacatcgc atcgagcgag cacgtactcg
3801 gatggaagcc ggtcttgtcg atcaggatga tctggacgaa gagcatcagg
3851 ggctcgcgcc agccgaactg ttcgccaggc tcaaggcgcg catgcccgac
3901 ggcgaggatc tcgtcgtgac ccatggcgat gcctgcttgc cgaatatcat
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4001 tggcggaccg ctatcaggac atagcgttgg ctaccctgta tattgctgaa
4051 gagcttggcg gcgaatgggc tgaccgcttc ctctgcttct acggtatcgc
4101 cgctcccgat tcgcagcgca tcgccttcta tcgccttctt gacgattct
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4351 caggacagca agggggagga ttgggaagac aatagcaggc atgctgggga
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5451 aactcacgtt aagggatttt ggtcatgaga ttatcaaaaa ggatcttcac
5501 ctagatcctt ttaaattaaa aatgaagttt taaatcaatc taaagtatat
5551 atgagtaaac ttggtctgac agttaccaat gcttaatcag tgaggcacct
5601 atctcagcga tctgtctatt tcgttcaccc atagttgcct gactccccgt
5651 cgtgtagata actacgatac gggagggctt accatctggc cccagtctg
5701 caatgatacc gcgagacca cgctcaccgg ctccagattt atcagcaata

```

FIGURE 16B

5751	aaccagccag	ccggaagggc	cgagcgcaga	agtggctcctg	caactttatc
5801	cgccctccatc	cagtctatta	attggtgccc	ggaagctaga	gtaagtagtt
5851	cgccaggttaa	tagtttgccg	aacggttggtg	ccattgctac	aggcatcgtg
5901	gtgtcacgct	cgtcggttgg	tatggcttca	ttcagctccg	gttcccaacg
5951	atcaaggcga	gttacatgat	cccccatggt	gtgcaaaaaa	gcggttagct
6001	ccttcgggtcc	tccgatcggt	gtcagaagta	agttggccgc	agtgttatca
6051	ctcatgggta	tggcagcact	gcataattct	cttactgtca	tgccatccgt
6101	aagatgcttt	tctgtgactg	gtgagtactc	aaccaagtca	ttctgagaat
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6201	accgcgccac	atagcagaac	tttaaaagtg	ctcatcattg	gaaaacgttc
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6401	aataagggcg	acacggaaat	gttgaatact	catactcttc	ctttttcaat
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6501	gaatgtattt	agaaaaataa	acaaataggg	gttccgcgca	catttccccg
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6601	ataaaaaatag	gcgtatcacg	aggcccttcc	gtctcgcgcg	tttcgggtgat
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6951	tcggcaaaat	cccttataaa	tcaaaagaat	agaccgagat	aggggtgagt
7001	gttgttccag	tttggaacaa	gagtcacta	ttaaagaacg	tggactccaa
7051	cgtcaaagg	cgaaaaaccg	tctatcaggg	cgatggccca	ctacgtgaac
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7201	gaacgtggcg	agaaaggaag	ggaagaaagc	gaaaggagcg	ggcgtagggg
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7401	ctgca				

FIGURE 16C

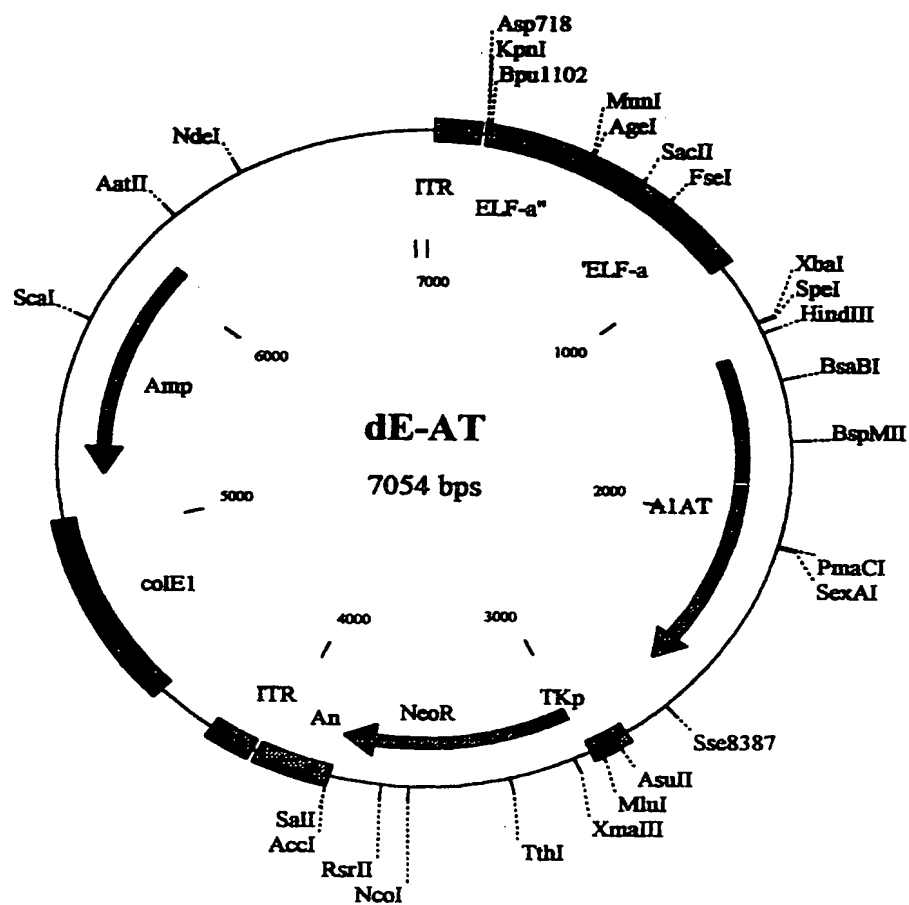


FIGURE 17

Molecule Name: dE-AT

7054 bps DNA Circular

Sequence Printed: 1-7054 (Full)

Date Printed 16 Apr 1999

Description: Fragment 2 Circularized

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301 ctctgcccctg ccttggggaa tcccaggagc cgtcggttaa ctcccactaa
351 cgtagaaccc agagatcgct gcgttcccgc cccctcaccg gcccgctctc
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2551 aatgattgaa caaaatacca agtctccctt ctcatggga aaagtggta
2601 atcccaccca aaaataactg cctctcgctc ctcaacctc cccctccatc
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2701 cccccccct gcagggggcc tgcagcagtg tggttttgca agaggaagca
```

FIGURE 17A

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2851	tttccacca	atgtcgagca	accccgccca	gcgtcttgtc	attggcgaat
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3251	tcgacgttgt	tcgacgaagc	ggaagggact	ggctgctatt	gggcgaagtg
3301	ccggggcagg	atctcctgtc	atctcacctt	gctcctgccc	agaaagtatc
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3401	gcccatcga	ccaccaagcg	aaacatcgca	tcgagcgagc	acgtactcgg
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FIGURE 17B

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6151	aatgtattta	gaaaaataaa	caaatagggg	ttccgcgcac	atttccccga
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6951	ttaatgcgcc	gctacagggc	gcgtcgcgcc	attcgccatt	caggctacgc
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7051	tgca				

FIGURE 17C

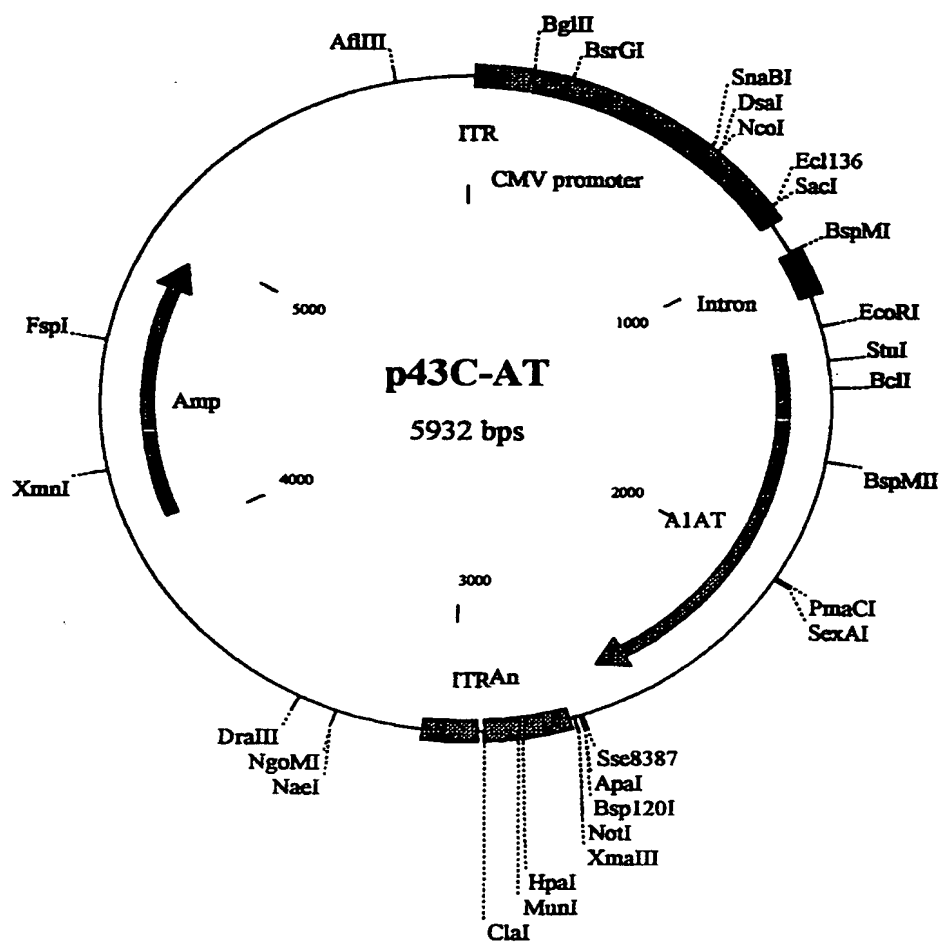


FIGURE 18

Molecule Name: p43C-AT 5932 bps DNA Circular
Sequence Printed: 1-5932 (Full) Date Printed 16 Apr 1999
Description: Ligation of TR and aat

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151 ctaggggttc ctatgtcttc aatattggcc attagccata ttattcattg
201 gttatatagc ataaatcaat attggctatt ggccattgca tacgttgat
251 ctatatcata atatgtacat ttatattggc tcatgtccaa tatgaccgcc
301 atgttggcat tgattattga ctagttatta atagtaatca attacggggt
351 cattagttca tagcccatat atggagttcc gcgttacata acttacggta
401 aatggccgcg ctggctgacc gcccaacgac ccccgcccat tgacgtcaat
451 aatgacgtat gttcccatag taacgccaat agggactttc cattgacgtc
501 aatgggtgga gtatttacgg taaactgccc acttggcagt acatcaagtg
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2651 ccccccccc ctgcaggggc cctcgaccgg ggccggcgct tcgagcagac
2701 atgataagat acattgatga gtttggacaa accacaacta gaatgcagt
```

FIGURE 18A

```

2751 aaaaaaatgc tttattttgtg aaattttgtga tgctatttgc ttattttgtaa
2801 ccattataag ctgcaataaaa caagttaaca acaacaattg cattcatttt
2851 atgttttcagg ttcaggggga gatgtgggag gtttttttaa gcaagtaaaa
2901 cctctacaaa tgtggtaaaa tcgataagga tcttaggaacc cctagtgatg
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```

FIGURE 18B

5751 ctttcctgcg ttatccccctg attctgtgga taaccgtatt accgcctttg
5801 agtgagctga taccgctcgc cgcagccgaa cgaccgagcg cagcgagtca
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5901 gcgttggccg attcattaat gcagggtgc ag

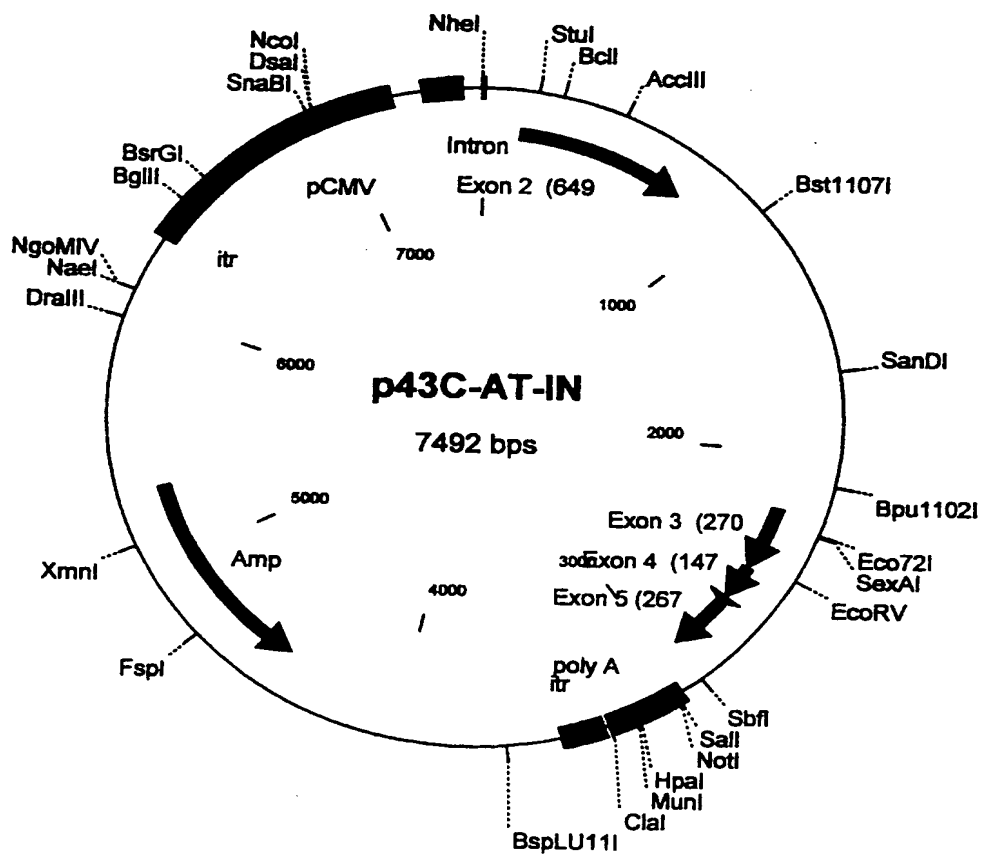


FIGURE 19

Molecule Name: p43C-AT-IN 7492 bps DNA Circular
 Sequence Printed: 1-7492 (Full) Date Printed 16 Apr 1999
 Description: Ligation of p43-C into IN

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101 ttgactgcct ggccccccca tctctgtcct gcaggacaat gccgtcttct
151 gtctcgtggg gcatcctcct gctggcaggc ctgtgctgcc tggctccctgt
201 ctccctggct gaggatcccc agggagatgc tgcccagaag acagatacat
251 cccaccatga tcaggatcac ccaaccttca acaagatcac ccccaacctg
301 gctgagttcg ccttcagcct ataccgccag ctggcacacc agtccaacag
351 caccaatatc ttcttctccc cagtgagcat cgctacagcc tttgcaatgc
401 tctccctggg gaccaaggct gacactcacg atgaaatcct ggagggcctg
451 aatttcaacc tcacggagat tccggaggct cagatccatg aaggcttcca
501 ggaactcctc cgtaccctca accagccaga cagccagctc cagctgacca
551 ccggcaatgg cctgttcctc agcagaggcc tgaagctagt ggataagttt
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701 gtactcaagg gaaaattgtg gatattggtca aggagcttga cagagacaca
751 gtttttgtct tggatgaatta catcttcttt aaaggtaagg ttgctcaacc
801 agcctgagct gtttcccata gaaacaagca aaaatatttc tcaaaccatc
851 agttcttgaa ctctccttgg caatgcatta tgggccatag caatgctttt
901 cagcgtggat tcttcagttt tctacacaca aacactaaaa tgttttccat
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1001 ctctgcagaa cttttcagag cttttaatgt ccttgtgtat actgtatatg
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1101 aagggatagt ttcattggaac atactttaca cgactctagt gtcccagaat
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2351 cactgtaaga agctgtccag ctgggtgctg ctgatgaaat acctgggcaa
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2551 tgatctgaag agcgtcctgg gtcaactggg catcactaag gtcttcagca
2601 atggggctga cctctccggg gtcacagagg aggcaccctc gaagctctcc
2651 aaggccgtgc ataaggctgt gctgaccatc gacgagaaag ggactgaagc
2701 tgctggggcc atgtttttag aggccatacc catgtctatc cccccgagg

```

FIGURE 19A

```

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2801 tctccctct tcatgggaaa agtgggtgaat cccacccaaa aataactgcc
2851 tctcgctcct caacccctcc cctccatccc tggcccccctc cctggatgac
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3051 gccgcttcga gcagacatga taagatacat tgatgagttt ggacaaacca
3101 caactagaat gcagtgaata aaatgcttta tttgtgaaat ttgtgagtct
3151 attgctttat ttgtaaccat tataagctgc aataaacaag ttaacaacaa
3201 caattgcatt cattttatgt ttcagggttca gggggagatg tgggaggttt
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3751 aaccgcacag gactataaag ataccaggcg tttccccctg gaagctccct
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5601 agcccgtcag ggcgcgtcag cgggtgttgg cgggtgtcgg ggctggctta
5651 actatcgggc atcagagcag attgtactga gagtgcacca tatgcggtgt
5701 gaaataccgc acagatgcgt aaggagaaaa taccgcatca ggaaattgta

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FIGURE 19B

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5751 aacgttaata ttttgttaaa attcgcgtta aatttttgtt aaatcagctc
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7351 aagactcttg cgtttctgat aggcacctat tgggtcttact gacatccact
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FIGURE 19C

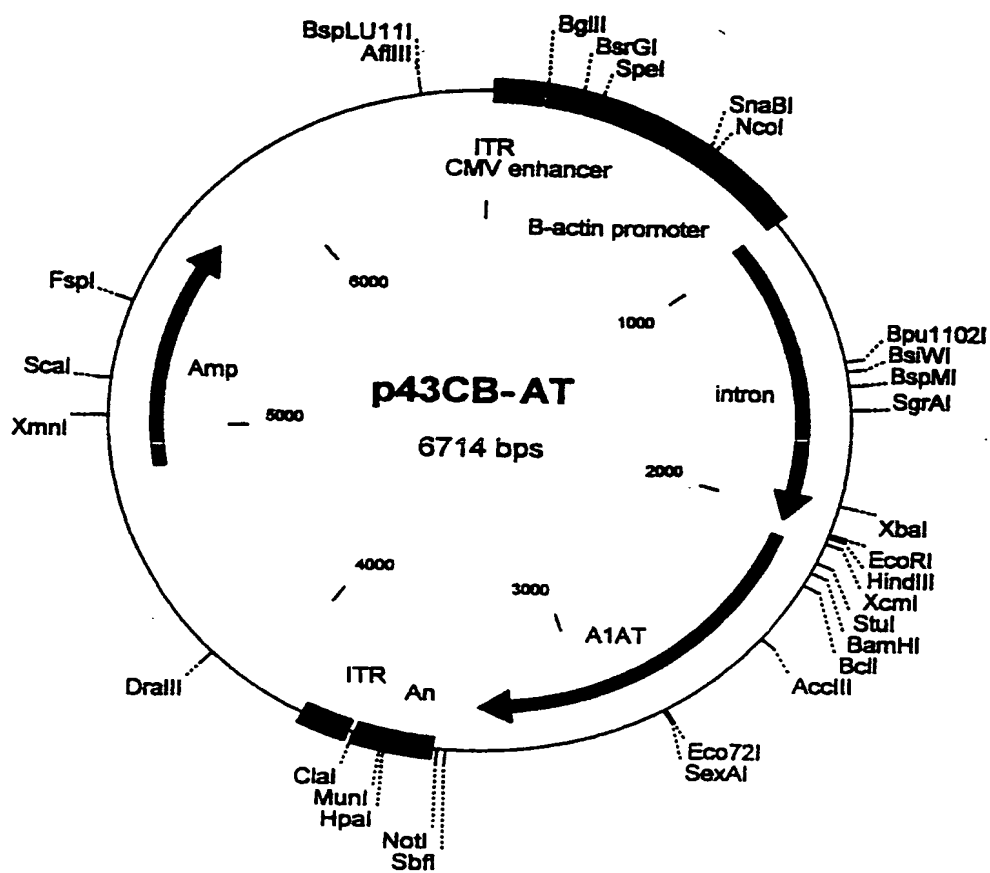


FIGURE 20

16 Apr 1999

Sequence Data

Page 1

Molecule: p43CB-AT, 6714 bps DNA Circular
Description: Ligation of Fragment 2 into Fragment 2
File Name: CB-AAT.cm5, dated 17 Nov 1998
Printed: 1-6714 bps (Full), format Single Strand

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151 ctaggggttc ctatgcttcc aatattggcc attagccata ttattcattg
201 gttatatagc ataaatcaat attggctatt ggccattgca tacgttgat
251 ctatatcata atatgtacat ttatattggc tcatgtccaa tatgaccgcc
301 atgttggcat tgattattga ctagtattta atagtaatca attacggggt
351 cattagttca tagcccatat atggagtacc gcgttacata acttacggta
401 aatggcccg ctaggtgacc gcccacgac ccccgcccat tgacgtcaat
451 aatgacgtat gttcccatag taacgccaat agggactttc cattgacgtc
501 aatgggtgga gtatttacgg taaactgccc acttggcagt acatcaagtg
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601 cgcctggcat tatgcccagt acatgacctt acgggacttt cctacttggc
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FIGURE 20A

p43CB-AT

Page 2

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2901	gggcaatgcc	accgccatct	tcttccctgcc	tgatgagggg	aaactacagc
2951	acctggaaaa	tgaactcacc	cacgatatca	tcaccaagtt	cctggaaaaat
3001	gaagacagaa	ggtctgccag	cttacattta	cccaaactgt	ccattactgg
3051	aacctatgat	ctgaagagcg	tcctgggtca	actgggcatc	actaagggtct
3101	tcagcaatgg	ggctgacctc	tccgggggtca	cagaggaggc	acccctgaag
3151	ctctccaagg	ccgtgcataa	ggctgtgctg	accatcgacg	agaaaaggac
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3251	ccgagggtcaa	gttcaacaaa	ccctttgtct	tcttaatgat	tgaacaaaaat
3301	accaagtctc	ccctcttcat	gggaaaagtg	gtgaatccca	cccaaaaaata
3351	actgcctctc	gctcctcaac	ccctccccctc	catccctggc	cccctccctg
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3451	gccctcgacc	cggcgccg	cttcgagcag	acatgataag	atacattgat
3501	gagtttggac	aaaccacaac	tagaatgcag	tgaaaaaaat	gctttatttg
3551	tgaattttgt	gatgctattg	ctttattttgt	aaccattata	agctgcaata
3601	aacaagttaa	caacaacaat	tgcatctcatt	ttatgtttca	ggttcagggg
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3701	aatcgataag	gatctaggaa	cccctagtga	tggagttggc	cactccctct
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3901	gaggcccgca	ccgatcgccc	ttcccaacag	ttgcgtagcc	tgaatggcga
3951	atggcgcgac	gcgccttgta	gcggcgcat	aagcgcgcg	ggtgtgggtg
4001	ttacgcgcag	cgtgaccgct	acacttgcca	gcgccttagc	gcccgtcct
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4101	agctctaaat	cgggggctcc	ctttaggggt	ccgatttagt	gctttacggc
4151	acctcgaccc	caaaaaactt	gattaggggtg	atggttcacg	tagtgggcca
4201	tcgccctgat	agacggtttt	tcgccctttg	acgttgaggt	ccacgttctt
4251	taatagtggg	ctcttggtcc	aaactggaac	aacactcaac	cctatctcgg
4301	tctattcttt	tgatttataa	gggattttgc	cgatttcggc	ctattgggtta
4351	aaaaatgagc	tgatttaaca	aaaattttaac	gcgaatttta	acaaaatatt
4401	aacgtttaca	atttcctgat	gcggtatttt	ctccttacgc	atctgtgcgg
4451	tatttcacac	cgcataatgg	gcactctcag	tacaatctgc	tctgatgccg
4501	catagttaag	ccagccccga	caccgcgcaa	caccgcgtga	cgcgccctga
4551	cgggcttgct	tgctcccggc	atccgcttac	agacaagctg	tgaccgtctc
4601	cgggagctgc	atgtgtcaga	ggtttttcacc	gtcatcaccg	aaacgcgcga
4651	gacgaaaggg	cctcgtgata	cgcctatttt	tataggttaa	tgtcatgata
4701	ataatggttt	cttagacgtc	aggtggcact	tttcggggaa	atgtgcgcgg
4751	aaccctatt	tgttttattt	tctaaataca	ttcaaataatg	tatccgctca
4801	tgagacaata	accctgataa	atgcttcaat	aatattgaaa	aaggaagagt
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4901	ttgccttcc	gtttttgctc	accagaaac	gctgggtgaaa	gtaaaagatg
4951	ctgaagatca	gttgggtgca	cgagtgggtt	acatcgaaact	ggatctcaac
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5051	gagcactttt	aaagtctctg	tatgtggcgc	ggtatttatcc	cgtattgacg
5101	ccgggcaaga	gcaactcggt	cgcgcatac	actattctca	gaatgacttg
5151	gttgagtact	caccagtcac	agaaaagcat	cttacggatg	gcatgacagt

FIGURE 20B

p43CB-AT

Page 3

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5251 acttacttct gacaacgatc ggaggaccga aggagctaac cgcttttttg
5301 cacaacatgg gggatcatgt aactcgcctt gatcgttggg aaccggagct
5351 gaatgaagcc ataccaaacg acgagcgtga caccacgatg cctgtagcaa
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5651 tatggatgaa cgaaatagac agatcgctga gataggtgcc tcaactgatta
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6201 acgggggggt cgtgcacaca gccagcttg gagegaacga cctacaccga
6251 actgagatac ctacagcgtg agcattgaga aagcgccacg cttcccgaag
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6701 atgcagggct gcag

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FIGURE 20C

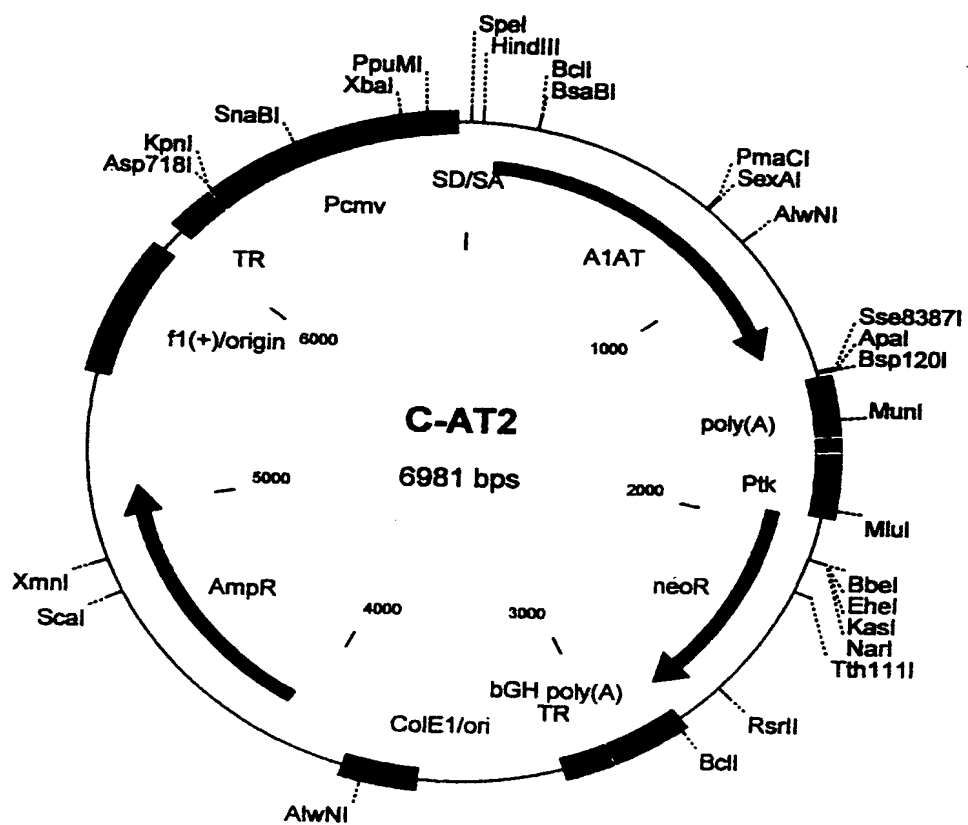


FIGURE 21

Molecule Name: C-AT2 6981 bps DNA Circular
 Sequence Printed: 1-6981 (Full) Date Printed 16 Apr 1999
 Description: Ligation of Fragment 1 and Fragment 2

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101 ttctgtctcgg tggggcatcc tcctgtctggc aggcctgtgc tgcctgggtcc
151 ctgtctccctt ggctgaggat ccccaggag atgctgcca gaagacagat
201 acatcccacc atgatcagga tcaccaacc ttcaacaaga tcaccccca
251 cctggctgag ttcgccttca gcctataccg ccagctggca caccagtcca
301 acagcaccaa tatcttcttc tccccagtga gcctcgctac agcctttgca
351 atgctctccc tggggaccaa ggctgacact cacgatgaaa tcctggaggg
401 cctgaatttc aacctcacgg agattccgga ggctcagatc catgaaggct
451 tccaggaact cctccgtacc ctcaaccagc cagacagcca gctccagctg
501 accaccggca atggcctggt cctcagcgag ggcctgaagc tagtggataa
551 gtttttggag gatgttaaaa agttgtacca ctcagaagcc ttcactgtca
601 acttcgggga caccgaagag gccaaagaaac agatcaacga ttacgtggag
651 aagggtactc aagggaataa tgtggatttg gtcaaggagc ttgacagaga
701 cacagttttt gctctgggtga attacatctt ctttaaaggc aaatgggaga
751 gaccttttga agtcaaggac accgaggaag aggacttcca cgtggaccag
801 gtgaccaccg tgaagggtgcc tatgatgaag cgtttaggca tgtttaacat
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901 gcaatgccac cgccatcttc ttcctgcctg atgaggggaa actacagcac
951 ctggaaaatg aactcaccca cgatatcatc accaagttcc tggaaaatga
1001 agacagaagg tctgccagct tacatttacc caaactgtcc attactggaa
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1251 gaggtcaagt tcaacaaacc ctttgtcttc ttaatgattg aacaaaatac
1301 caagtctccc ctcttcattg gaaaagtggg gaatcccacc caaaaataac
1351 tgctctcgc tcctcaacc ccccttcca tccctggccc cctccctgga
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1501 acaaaccaca actagaatgc agtgaaaaaa atgctttatt tgtgaaattt
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1651 ggaggttttt tagtcgacct cgagcagtggt ggttttgcaa gaggaagcaa
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1751 gtggttttgc aagaggaagc aaaaagcctc tccaccagg cctggaatgt
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1901 attaaggtga cgcgtgtggc ctcgaaacac gagcgacct gcagccaata
1951 tgggatcggc cattgaacaa gatggattgc acgcaggttc tccggccgct
2001 tgggtggaga ggctattcgg ctatgactgg gcacaacaga caatcggctg
2051 ctctgatgcc gccgtgttcc ggctgtcagc gcaggggccc ccggttcttt
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2251 cggggcagga tctcctgtca tctcaccttg ctccctgcga gaaagtatcc
2301 atcatggctg atgcaatgcg gcggctgcat acgcttgatc cggctacctg
2351 cccattcgac caccaagcga aacatcgcat cgagcgagca cgtactcgga
2401 tggaaagcgg tcttgtcgat caggatgatc tggacgaaga gcatcagggg
2451 ctgcgcccag ccgaactgtt cgccaggctc aaggcgcgca tgcccagcgg
2501 cgaggatctc gtcgtgaccc atggcgatgc ctgcttgccg aatatcatgg
2551 tggaaaatgg ccgcttttct ggattcatcg actgtggccc gctgggtgtg
2601 gcggaccgct atcaggacat agcgttggct acccgtgata ttgctgaaga
2651 gcttggcggc gaatgggctg accgcttctt cgtgctttac ggtatcgccg
2701 ctcccgatcc gcagcgcac gccttctatc gccttcttga cgagttcttc

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FIGURE 21A

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2851	tggaaagggtg	cactcccact	gtcctttcct	aataaaatga	ggaaattgca
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2951	ggacagcaag	ggggaggatt	gggaagacaa	tagcaggcat	gctggggaga
3001	gatctaggaa	cccctagtga	tggagttggc	cactccctct	ctgcgcgctc
3051	gctcgctcac	tgaggccgcc	cgggcaaagc	ccgggcgtcg	ggcgaccttt
3101	ggtcgccccg	cctcagttag	cgagcgagcg	cgcagagagg	gagtggccaa
3151	ccccccccc	ccccccctg	cagccctgca	ttaatgaatc	ggccaacgcg
3201	cggggagagg	cgggtttgct	attgggctg	cttcgcttc	ctcgctcact
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3351	acatgtgagc	aaaaggccag	caaaaggcca	ggaaccgtaa	aaaggccgcg
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4051	ctcacgttaa	gggatttttg	tcattgagatt	atcaaaaagg	atcttcacct
4101	agatcctttt	aaattaaaaa	tgaagtttta	aatcaatcta	aagtatatat
4151	gagtaaactt	ggtctgacag	ttaccaatgc	ttaatcagtg	aggcacctat
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4251	tgtagataac	tacgatacgg	gagggcttac	catctggccc	cagtgtcgca
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4651	catggttatg	gcagcactgc	ataattctct	tactgtcatg	ccatccgtaa
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4901	taaccctact	gtgcacccaa	ctgatcttca	gcattctttaa	ctttcaccag
4951	cgtttctggg	tgagcaaaaa	caggaaggca	aaatgccgca	aaaaagggaa
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5101	atgtatttag	aaaaataaac	aaataggggt	tccgcgcaca	tttccccgaa
5151	aagtgccacc	tgacgtctaa	gaaaccatta	ttatcatgac	attaacctat
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5251	cggtgaaaaa	ctctgacaca	tgcagctccc	ggagacgggc	acagcttggtc
5301	tgtaagcggg	tgcggggagc	agacaagccc	gtcagggcgc	gtcagcgggt
5351	gttggcgggt	gtcggggctg	gcttaactat	gcggcatcag	agcagattgt
5401	actgagagtg	caccatattgc	ggtgtgaaat	accgcacaga	tgcgtaaggga
5451	gaaaataccg	catcaggaaa	ttgtaaacgt	taatattttg	ttaaaattcg
5501	cgttaaattt	ttgttaaata	agctcatttt	ttaaaccaata	ggccgaaatc
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5601	tgttccagtt	tggaacaaga	gtccactatt	aaagaacgtg	gactccaacg
5651	tcaaagggcg	aaaaaccgtc	tatcagggcg	atggccact	acgtgaacca
5701	tcaccctaata	caagtttttt	ggggctcgagg	tgccgtaaaag	cactaaatcg

FIGURE 21B

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5751 gaaccctaaa gggagccccc gatttagagc ttgacgggga aagccggcga
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5851 ctggcaagtg tagcggtcac gctgcgcgta accaccacac ccgccgcgct
5901 taatgcgccg ctacagggcg cgtcgcgcca ttcgccattc aggctacgca
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FIGURE 21C

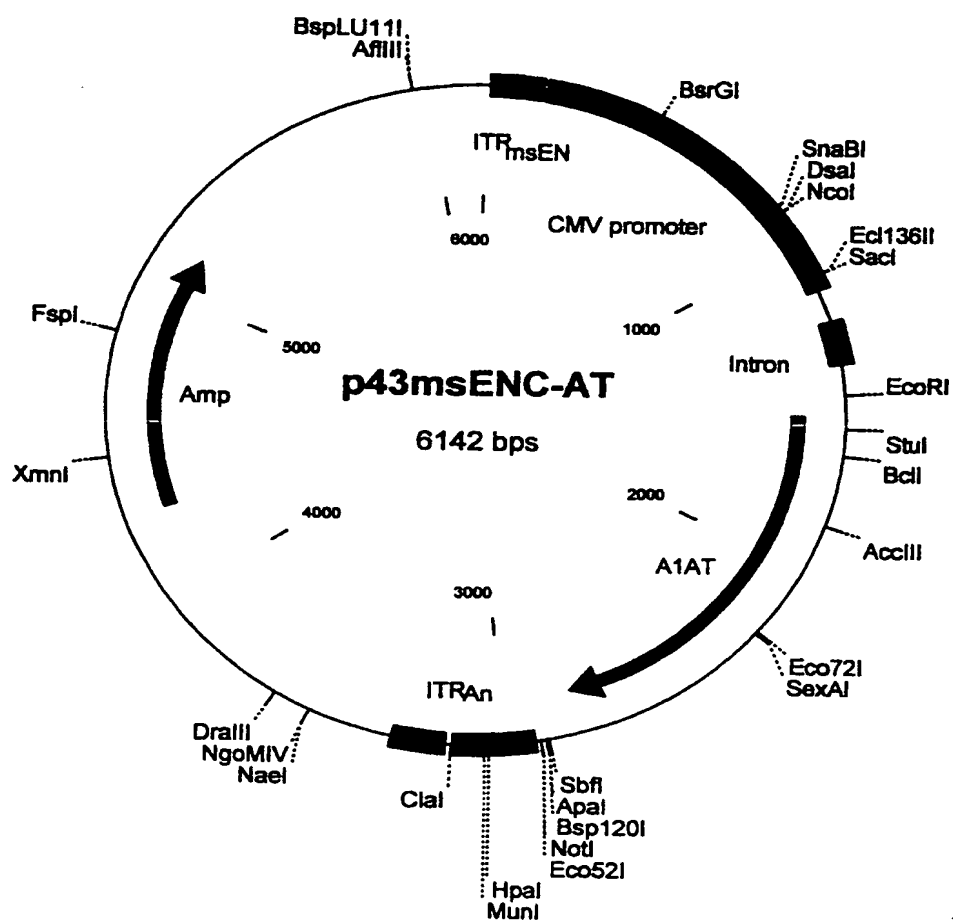


FIGURE 22

19 Apr 1999

Sequence Data

Page 1

Molecule: p43msENC-AT, 6142 bps DNA Circular
Description: Ligation of inverted msEnhancer into p43-AAT*
File Name: p43smENC-AT.cm5, dated 19 Apr 1999
Printed: 1-6142 bps (Full), format Single Strand

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151 ctaggggttc ctatgcttga caccacaata tggcctgggg tgaggaatgg
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251 ctcccgggag ttatttttag agcgccaaca cctgctgcct gccaccattt
301 cctcaccgct ctaaaaataa ctcccacca ttcctcaccg gtcgccatat
351 ttgggtgtcg tgaggaatgg tgagatcttc aatattggcc attagccata
401 ttattcattg gttatatagc ataaatcaat attggctatt ggccattgca
451 tacgttgtat ctatatcata atatgtacat ttatattggc tcatgtccaa
501 tatgaccgcc atgttggcat tgattattga ctagtatta atagtaatca
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1101 gtttagtgaa ccgtcagatc actagaagct ttattgcggt agtttatcac
1151 agtttaaattg ctaacgcagt cagtgttctt gacacaacag tctcgaactt
1201 aagctgcaga agttggtcgt gaggcactgg gcaggtaagt atcaaggtta
1251 caagacaggt ttaaggagac caatagaaac tgggcttggt gagacagaga
1301 agactcttgc gtttctgata ggcacctatt ggtcttactg acatccactt
1351 tgcccttctc tccacaggtg tccactccca gttcaattac agctcttaag
1401 gctagagtac ttaatacgac tcaactatagg ctagaactag tggatcccc
1451 gggctgcagg aattcgatat caagcttggg gattttcagg caccaccact
1501 gacctgggac agtgaatcga caatgccgtc ttctgtctcg tggggcatcc
1551 tctgctggc aggcctgtgc tgccgtgtcc ctgtctccct ggctgaggat
1601 cccaggggag atgctgcccga gaagacagat acatcccacc atgatcagga
1651 tcacccaacc ttcaacaaga tcaccccaaa cctggctgag ttgccttca
1701 gcctataacc ccagctggca caccagtcca acagcaccaa tatcttcttc
1751 tccccagtga gcatcgctac agcctttgca atgctctccc tggggaccaa
1801 ggctgacact cacgatgaaa tcttgagggg cctgaatttc aacctcacgg
1851 agattccgga ggctcagatc catgaaggct tccaggaact cctccgtacc
1901 ctcaaccagc cagacagcca gctccagctg accaccggca atggcctgtt
1951 cctcagcgag ggcctgaagc tagtgataaa gtttttggag gatgttaaaa
2001 agttgtacca ctcaagaagc ttcactgtca acttcgggga caccgaagag
2051 gccaaagaac agatcaacga ttacgtggag aagggtaactc aagggaaaat
2101 tgtggatttg gtcaaggagc ttgacagaga cacagttttt gctctggtga
2151 attacatctt ctttaaaggc aaatgggaga gaccctttga agtcaaggac
2201 accgaggaag aggacttcca cgtggaccag gtgaccaccg tgaagggtgc
2251 tatgatgaag cgtttaggga tgtttaacat ccagcactgt aagaagctgt
2301 ccagctgggt gctgctgatg aaatacctgg gcaatgccac cgccatcttc
2351 ttcctgcctg atgaggggaa actacagcac ctggaaaatg aactcaccca
2401 cgatatcatc accaagttcc tggaaaatga agacagaagg tctgccagct
```

FIGURE 22A

p43msENC-AT

Page 2

```
2451  tacatttacc caaactgtcc attactggaa cctatgatct gaagagcgtc
2501  ctgggtcaac tgggcatcac taaggctctc agcaatgggg ctgacctctc
2551  cggggtcaca gaggaggcac ccctgaagct ctccaaggcc gtgcataagg
2601  ctgtgctgac catcgacgag aaagggactg aagctgctgg ggccatgttt
2651  ttagaggcca tacccatgtc tatccccccc gaggtcaagt tcaacaaacc
2701  ctttgtcttc ttaatgattg aacaaaatac caagtctccc ctcttcatgg
2751  gaaaagtggg gaatcccacc caaaaataac tgccctctcg tcctcaacc
2801  ctcccctcca tccctggccc cctccctgga tgacattaaa gaagggttga
2851  gctggtaacc cccccccccc ctgcaggggc cctcgacccg ggcgccgct
2901  tcgagcagac atgataagat acattgatga gtttggacaa accacaacta
2951  gaatgcagtg aaaaaaatgc tttatttgtg aaatttgtga tgctattgct
3001  ttatttgtaa ccattataag ctgcaataaa caagttaaca acaacaattg
3051  cattcatttt atgtttcagg ttcaggggga gatgtgggag gttttttaa
3101  gcaagtaaaa cctctacaaa tgtggtaaaa tcgataagga tctaggaacc
3151  cctagtgatg gagttggcca ctccctctct gcgcgctcgc tcgctcactg
3201  aggccgcccc ggcaaaagccc gggcgctcgg cgacctttgg tcgccccggc
3251  tcagtgagcg agcgagcgcg cagagaggga gtggccaacc cccccccccc
3301  cccccctgca gcctggcgta atagcgaaga ggcccgccac gatcgccctt
3351  cccaacagtt gcgtagcctg aatggcggaat ggcgcgacgc gccctgtagc
3401  ggcgcattaa gcgcggcggg tgtggtggtt acgcgcagcg tgaccgtac
3451  acttgccagc gccctagcgc ccgctccttt cgctttcttc ccttcttttc
3501  tcgccacggt cgccggcttt ccccgctcaag ctctaaatcg ggggctccct
3551  ttagggttcc gatttagtgc tttacggcac ctcgacccca aaaaacttga
3601  ttaggggtgat ggttcacgta gtgggccatc gccctgatag acggtttttc
3651  gccctttgac gttggagtcc acgttcttta atagtggact cttgttccaa
3701  actggaacaa cactcaaccc tatctcggtc tattcttttg atttataagg
3751  gattttgccc atttcggcct attgggttaa aaatgagctg atttaacaaa
3801  aatttaacgc gaattttaac aaaatattaa cgtttacaat ttcctgatgc
3851  ggtattttct ccttacgcac ctgtgcggtg tttcacaccg catatggtgc
3901  actctcagta caatctgctc tgatgccgca tagttaagcc agccccgaca
3951  cccgccaaaca cccgctgacg cgccctgacg ggcttgtctg ctcccgcgat
4001  ccgcttacag acaagctgtg accgtctccg ggagctgcat gtgtcagagg
4051  ttttcaccgt catcaccgaa acgcgcgaga cgaaagggcc tcgtgatagc
4101  cctattttta taggttaatg tcatgataat aatggtttct tagacgtcag
4151  gtggcacttt tcggggaaat tcgcgcggaa cccctatttg tttatttttc
4201  taaatacatt caaatatgta tccgctcatg agacaataac cctgataaat
4251  gcttcaataa tattgaaaaa ggaagagtat gagtattcaa catttccgtg
4301  tcgcccttat tccctttttt gcggcatttt gccttcctgt ttttgctcac
4351  ccagaaacgc tggtgaaagt aaaagatgct gaagatcagt tgggtgcacg
4401  agtgggttac atcgaactgg atctcaacag cggtaaagatc cttgagagtt
4451  ttcgccccga agaacgtttt ccaatgatga gcacttttaa agttctgcta
4501  tgtggcgcgg tattatcccg tattgacgcc gggcaagagc aactcggtcg
4551  ccgcatacac tattctcaga atgacttggg tgagtactca ccagtcacag
4601  aaaagcatct tacggatggc atgacagtaa gagaattatg cagtgtgcc
4651  ataaccatga gtgataaac tgccggccaac ttacttctga caacgatcgg
4701  aggaccgaag gagctaaccg cttttttgca caacatgggg gatcatgtaa
4751  ctgcgcttga tcgttgggaa ccggagctga atgaagccat accaaacgac
4801  gagcgtgaca ccacgatgcc tgtagcaatg gcaacaacgt tgcgcaact
4851  attaaactgg gaactactta ctctagcttc ccggcaacaa ttaatagact
4901  ggatggaggc ggataaagtt gcaggaccac ttctgcgctc ggcccttccg
4951  gctggctggt ttattgctga taaacttgga gccggtgagc gtgggtctcg
5001  cggtatcatt gcagcactgg gcccagatgg taagccctcc cgtatcgtag
5051  ttatctacac gacggggagt caggcaacta tggatgaacg aaatagacag
5101  atcgctgaga taggtgcctc actgattaag cattggtaac tgtcagacca
5151  agtttactca tatatacttt agattgattt aaaacttcat ttttaattta
```

FIGURE 22B

p43msENC-AT

Page 3

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5201 aaaggatcta ggtgaagatc ctttttgata atctcatgac caaaatccct
5251 taacgtgagt ttctggtcca ctgagcgtca gaccccgtag aaaagatcaa
5301 aggatcttct tgagatcctt tttttctgcg cgtaatctgc tgcttgcaaa
5351 caaaaaaacc accgctacca gcggtgggtt gtttgccgga tcaagagcta
5401 ccaactcttt ttccgaaggt aactggcttc agcagagcgc agataccaaa
5451 tactgtcctt ctagtgtagc cgtagttagg ccaccacttc aagaactctg
5501 tagcaccgcc tacatacctc gctctgctaa tcctgttacc agtggctgct
5551 gccagtggcg ataagtcgtg tcttaccggg ttggactcaa gacgatagtt
5601 accggataag gcgcagcggg cgggctgaac ggggggttcg tgcacacagc
5651 ccagcttgga gcgaacgacc tacaccgaac tgagatacct acagcgtgag
5701 cattgagaaa gcgccacgct tcccgaaggg agaaaggcgg acaggtatcc
5751 ggtaagcggc agggtcggaa caggagagcg cacgagggag cttccagggg
5801 gaaacgcctg gtatctttat agtcctgtcg ggtttcgcca cctctgactt
5851 gagcgtcgat ttttgtgatg ctcgtcaggg gggcggagcc tatggaaaaa
5901 cgccagcaac gcggcctttt tacggttcct ggccttttgc tggccttttg
5951 ctcacatggt ctttcctgcg ttatcccctg attctgtgga taaccgtatt
6001 accgcctttg agtgagctga taccgctcgc cgcagccgaa cgaccgagcg
6051 cagcgagtca gtgagcgagg aagcggaaga gcgcccaata cgcaaaccgc
6101 ctctccccgc gcgttggccg attcattaat gcagggctgc ag
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FIGURE 22C

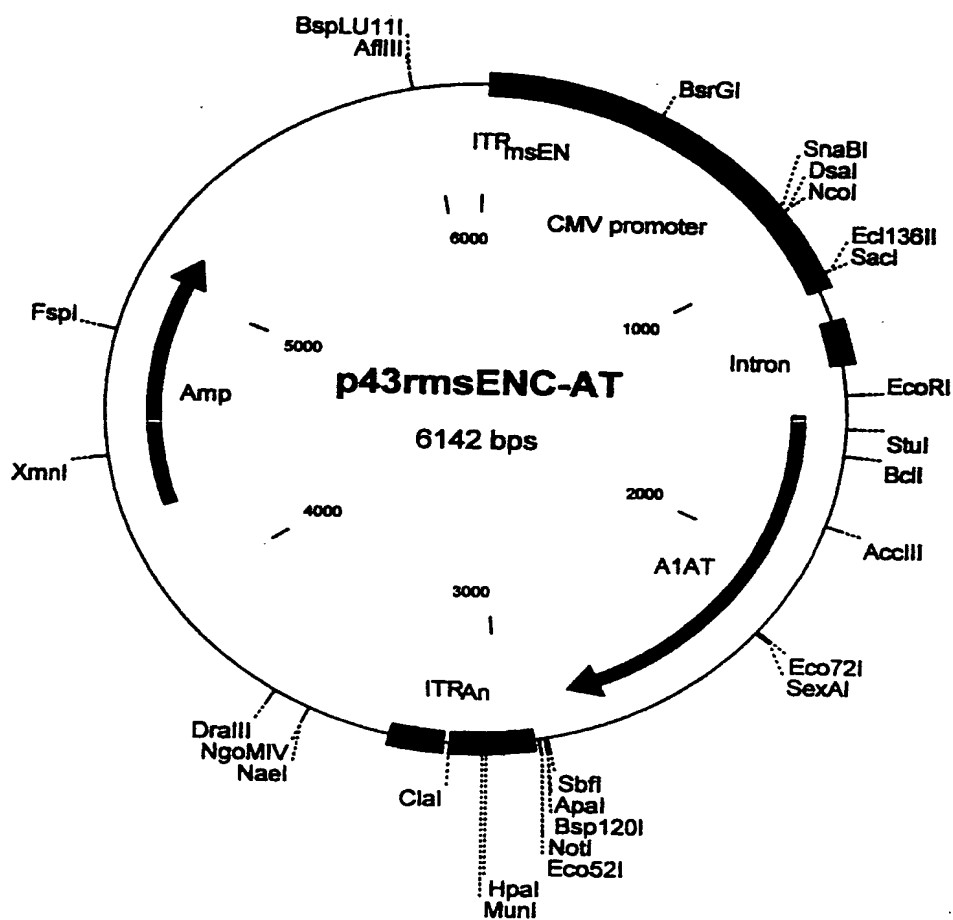


FIGURE 23

19 Apr 1999

Sequence Data

Page 1

Molecule: p43rmsENC-AT, 6142 bps DNA Circular
Description: Ligation of inverted msEnhancer into p43-AAT*
File Name: p43rmsENC-AT.cm5, dated 19 Apr 1999
Printed: 1-6142 bps (Full), format Single Strand

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1  gggggggggg ggggggggttg gccactccct ctctgcgcgc tcgctcgctc
51  actgaggccg ggcgaccaa ggtcgcccga cgcccgggct ttgcccgggc
101 ggcctcagtg agcgagcgag cgcgagaga gggagtggcc aactccatca
151 ctaggggttc ctatgctga caccacaata tggcctgggg tgaggaatgg
201 tgccgtcgcc atatttgggt gtccaccatt cctcaccgct ctaaaaataa
251 ctcccgggag ttatttttag agcgccaaca cctgctgcct gccaccatt
301 cctcaccgct ctaaaaataa ctcccacca ttcctcacc gtcgccatat
351 ttgggtgtcg tgaggaatgg tgagatcttc aatattggcc attagccata
401 ttattcattg gttatatagc ataaatcaat attggctatt ggccattgca
451 tacgttgtat ctatatcata atatgtacat ttatatggc tcatatgcaa
501 tatgaccgcc atgttggcat tgattattga ctagtatta atagtaatca
551 attacggggt cattagttca tagcccatat atggagtcc gcgttacata
601 acttacggta aatggccgc ctggctgacc gcccaacgac ccccgcccat
651 tgacgtcaat aatgacgtat gtcccatag taacgccaat agggactttc
701 cattgacgtc aatgggtgga gtatttacgg taaactgccc acttggcagt
751 acatcaagt tatcatatgc caagtccgcc ccctattgac gtcaatgacg
801 gtaaatggcc cgcctggcat tatgccagc acatgacctt acgggacttt
851 cctacttggc agtacatcta cgtattagtc atcgctatta ccatgggtgat
901 gcggttttgg cagtacacca atggcgctgg atagcggtt gactcacggg
951 gatttccaag tctccacccc attgacgtca atgggagttt gttttggcac
1001 caaaatcaac gggactttcc aaaatgtcgt aataacccc cccggttgac
1051 gcaaattggc ggtagcgctg tacggtggga ggtctatata agcagagctc
1101 gtttagtgaa ccgtcagatc actagaagct ttattgcggt agtttatcac
1151 agttaaattg ctaacgcagt cagtgttct gacacaacag tctcgaactt
1201 aagctgcaga agttggtcgt gaggcactgg gcaggtaagt atcaaggtta
1251 caagacaggt ttaaggagac caatagaaac tgggcttgct gagacagaga
1301 agactcttgc gtttctgata ggcacctatt ggtcttactg acatccactt
1351 tgcctttctc tccacaggtg tccactccca gttcaattac agctcttaag
1401 gctagagtac ttaatacgac tcactatagg ctagaactag tggatcccc
1451 gggctgcagg aattcgatat caagcttggg gattttcagg caccaccact
1501 gacctgggac agtgaatcga caatgccgtc ttctgtctcg tggggcatcc
1551 tcctgctggc aggcctgtgc tgccgtgtcc ctgtctccct ggctgaggat
1601 ccccagggag atgctgcca gaagacagat acatcccacc atgatcagga
1651 tcacccaacc ttcaacaaga tcacccccaa cctggctgag ttcgccttca
1701 gcctataacc ccagctggca caccagtcca acagcaccia tatcttcttc
1751 tcccagtgga gcatcgctac agcctttgca atgctctccc tggggaccaa
1801 ggctgacact cagcatgaaa tcctggaggg cctgaatttc aacctcacgg
1851 agattccgga ggctcagatc catgaaggct tccaggaact cctccgtacc
1901 ctcaaccagc cagacagcca gctccagctg accaccggca atggcctgtt
1951 cctcagcgag ggcctgaagc tagtggataa gtttttggag gatgttaaaa
2001 agttgtacca ctcagaagcc ttcactgtca acttcgggga caccgaagag
2051 gccaagaaac agatcaacga ttacgtggag aagggtactc aagggaatat
2101 tgtggatttg gtcaaggagc ttgacagaga cacagttttt gctctggtga
2151 attacatctt ctttaaaggc aaatgggaga gaccctttga agtcaaggac
2201 accgaggaag aggacttcca cgtggaccag gtgaccaccg tgaagggtgc
2251 tatgatgaag cgtttaggca ctgttaacat ccagcactgt aagaagctgt
2301 ccagctgggt gctgctgatg aaatacctgg gcaatgccac cgccatcttc
2351 ttcctgcctg atgaggggaa actacagcac ctggaaaatg aactcaccca
2401 cgatatcatc accaagttcc tggaaaatga agacagaagg tctgccagct
```

FIGURE 23A

p43rmsENC-AT

Page 2

```

2451   tacattttacc caaactgtcc attactggaa cctatgatct gaagagcgtc
2501   ctgggtcaac tgggcatcac taaggctctc agcaatgggg ctgacctctc
2551   cggggtcaca gaggaggcac ccctgaagct ctccaaggcc gtgcataagg
2601   ctgtgctgac catcgacgag aaagggactg aagctgctgg ggccatgttt
2651   ttagaggcca taccatgtc tatccccccc gaggtcaagt tcaacaaacc
2701   ctttgtcttc ttaatgattg aacaaaatac caagtctccc ctcttcattg
2751   gaaaagtggg gaatcccacc caaaaataac tgcctctcgc tcctcaaccc
2801   ctcccccca tccctggccc cctccctgga tgacattaaa gaagggttga
2851   gctggtaacc cccccccccc ctgcaggggc cctcgaccgc ggccggccgt
2901   tcgagcagac atgataagat acattgatga gtttggacaa accacaacta
2951   gaatgcagtg aaaaaaatgc tttatttgtg aaatttgtga tgctattgct
3001   ttatttgtaa ccattataag ctgcaataaa caagttaaca acaacaattg
3051   cattcatttt atgtttcagg ttcaggggga gatgtgggag gttttttaa
3101   gcaagtaaaa cctctacaaa tgtgttaaaa tcgataagga tctaggaacc
3151   cctagtgatg gagttggcca ctccctctct gcgcgctcgc tcgctcactg
3201   aggccgcccc ggcaaagccc gggcgctcgg cgacctttgg tcgcccggcc
3251   tcagtgagcg agcgagcgcg cagagaggga gtggccaacc cccccccccc
3301   cccccctgca gcctggcgta atagcgaaga ggcccgcacc gatcgccctt
3351   cccaacagtt gcgtagcctg aatggcgaat ggcgcgacgc gccctgtagc
3401   ggcgcattaa gcgcggcggg tgtgggtggt acgcgcagcg tgaccgtac
3451   acttgccagc gccctagcgc ccgctccttt cgctttcttc ccttcctttc
3501   tcgccacgtt cgccggcctt ccccgtaacg ctctaaatcg ggggctccct
3551   ttagggttcc gatttagtgc tttacggcac ctcgacccca aaaaacttga
3601   ttagggtgat ggttcacgta gtgggccatc gccctgatag acgggttttc
3651   gccctttgac gttggagtcc acgttcttta atagtggact cttgttccaa
3701   actggaacaa cactcaaccc tatctcggtc tattcttttg atttataagg
3751   gattttgccg atttcggcct attgggttaa aaatgagctg atttaacaaa
3801   aatttaacgc gaattttaac aaaatattaa cgtttacaat ttctgatgc
3851   ggtaattttc ccttacgcac ctgtgcggta tttcacaccg catatggtgc
3901   actctcagta caatctgctc tgatccgca tagttaagcc agccccgaca
3951   cccgccaaca cccgctgacg cgccctgacg ggcttgtctg ctcccgcat
4001   ccgcttacag acaagctgtg accgtctccg ggagctgcat gtgtcagagg
4051   ttttcaccgt catcaccgaa acgcgcgaga cgaaagggcc tcgtgatacg
4101   cctattttta taggttaatg tcatgataat aatggtttct tagacgtcag
4151   gtggcacttt tcggggaatg gtgcgcggaa cccctatttg tttatttttc
4201   taaatacatt caaatatgta tccgctcatg agacaataac cctgataaat
4251   gcttcaataa tattgaaaaa ggaagagtat gagtattcaa catttcctg
4301   tcgcccttat tccctttttt gcggcatttt gccttcctgt ttttgcac
4351   ccagaaacgc tgggtgaaagt aaaagatgct gaagatcagt tgggtgcacg
4401   agtgggttac atcgaaactg atctcaacag cggttaagatc cttgagagtt
4451   ttcgccccga agaacgtttt ccaatgatga gcacttttaa agttctgcta
4501   tgtggcgcgg tattatcccg tattgacgcc gggcaagagc aactcggtcg
4551   ccgcatacac tattctcaga atgacttggg tgagtactca ccagtcacag
4601   aaaagcatct tacggatggc atgacagtaa gagaattatg cagtgtgcc
4651   ataaccatga gtgataacac tgcggccaac ttacttctga caacgatcgg
4701   aggaccgaag gagctaaccg cttttttgca caacatgggg gatcatgtaa
4751   ctgcgcttga tcggtgggaa ccggagctga atgaagccat accaaacgac
4801   gagcgtgaca ccacgatgcc tgtagcaatg gcaacaacgt tgcgcaaact
4851   attaaactggc gaactactta ctctagcttc ccggcaacaa ttaatagact
4901   ggatggaggc ggataaagtt gcaggaccac ttctgcgctc ggcccttcgg
4951   gctggctggg ttattgctga taaatctgga gccggtgagc gtgggtctcg
5001   cggtatcatt gcagcactgg ggcagatgg taagccctcc cgtatcgtag
5051   ttatctacac gacggggagt caggcaacta tggatgaacg aaatagacag
5101   atcgctgaga taggtgcctc actgattaag cattggtaac tgtcagacca
5151   agtttactca tatatacttc agattgattt aaaacttcat ttttaattta

```

FIGURE 23B

p43rmsENC-AT

Page 3

```
5201 aaaggatcta ggtgaagatc ctttttgata atctcatgac caaaatccct
5251 taacgtgagt ttctgttcca ctgagcgtca gaccccgtag aaaagatcaa
5301 aggatcttct tgagatcctt tttttctgcg cgtaatctgc tgcttgcaaa
5351 caaaaaaacc accgctacca gcggtggttt gtttgccgga tcaagagcta
5401 ccaactcttt ttccgaaggt aactggcttc agcagagcgc agatacaaaa
5451 tactgtcctt ctagtgtagc cgtagttagg ccaccacttc aagaactctg
5501 tagcaccgcc tacatacctc gctctgctaa tcctgttacc agtggctgct
5551 gccagtggcg ataagtcgtg tcttaccggg ttggactcaa gacgatagtt
5601 accggataag gcgcagcggc cgggctgaac ggggggttcg tgcacacagc
5651 ccagcttgga gcgaacgacc tacaccgaac tgagatacct acagcgtgag
5701 cattgagaaa gcgccacgct tcccgaaggg agaaaggcgg acaggtatcc
5751 ggtaagcggc agggtcggaa caggagagcg cacgagggag cttccagggg
5801 gaaacgcctg gtatctttat agtcctgtcg ggtttcgcca cctctgactt
5851 gagcgtcgat ttttgtgatg ctcgtcaggg gggcggagcc tatggaaaaa
5901 cgccagcaac gcggcctttt tacggttcct ggccttttgc tggccttttg
5951 ctcacatggt ctttcctgcg ttatccccctg attctgtgga taaccgtatt
6001 accgcctttg agtgagctga taccgctcgc cgcagccgaa cgaccgagcg
6051 cagcgagtca gtgagcgagg aagcggaaga gcgccaata cgcaaaccgc
6101 ctctccccgc gcgttggccg attcattaat gcagggtgc ag
```

FIGURE 23C

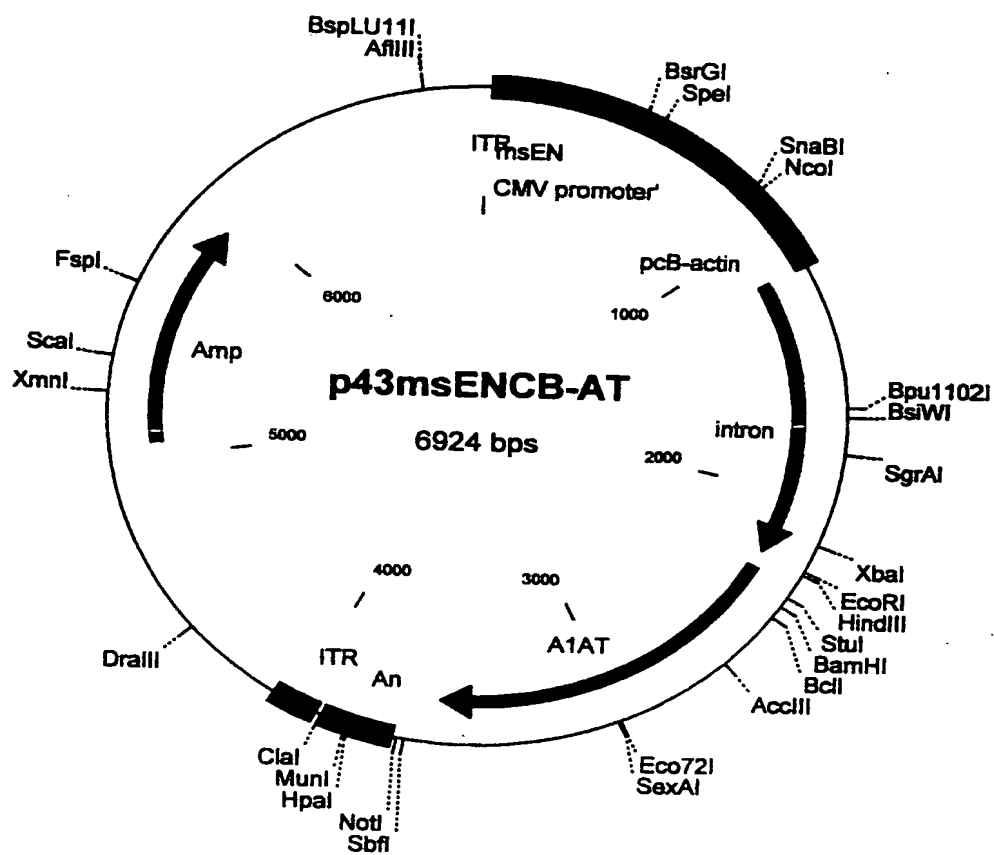


FIGURE 24

19 Apr 1999

Sequence Data

Page 1

Molecule: p43msENCB-AT, 6924 bps DNA Circular
Description: Ligation of msEnhacer into p43CB-AT*
File Name: p43msENCB-AT.cm5, dated 19 Apr 1999
Printed: 1-6924 bps (Full), format Single Strand

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101 ggcctcagtg agcgagcgag cgcgagaga gggagtggcc aactccatca
151 ctaggggttc ctagatctca ccattcctca cgacacccaa atatggcgac
201 ggggtgaggaa tgggtggggag ttatttttag agcgggtgagg aatgggtgggc
251 aggcagcagg tgttggcgct ctaaaaataa ctcccgggag ttatttttag
301 agcgggtgagg aatgggtggac acccaaatat ggcgacggca ccattcctca
351 ccccaggcca tatttgggtg tcagatcttc aatattggcc attagccata
401 ttattcattg gttatatagc ataaatcaat attggctatt ggccattgca
451 tacgttgtat ctatatcata atatgtacat ttatattggc tcatatgcaa
501 tatgaccgcc atgttggcat tgattattga ctagtatta atagtaatca
551 attacggggt cattagttca tagcccatat atggagtacc gcgttacata
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701 cattgacgtc aatgggtgga gtatttacgg taaactgccc acttggcagt
751 acatcaagtg tatcatatgc caagtccgcc ccctattgac gtcaatgacg
801 gtaaatggcc cgcctggcat tatgccagat acatgacctt acgggacttt
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901 ggtgagcccc acgttctgct tcactctccc catctccccc ccctcccccac
951 cccaattttt gtatttattt attttttaat tattttgtgc agcgatgggg
1001 gcgggggggg gggggggggcg cgcgccaggg ggggcggggc gggcgagggg
1051 gcggggcggg gcgagggcgga gaggtgcggc ggcagccaat cagagcgggc
1101 cgctccgaaa gtttcctttt atggcgaggg ggcggcgggc gcggccctat
1151 aaaaagcgaa gcgcgcggcg ggcgggagtc gctgcgacgc tgccttcgcc
1201 ccgtgccccg ctccgcccgc gctcgcgcgc gcccgccccg gctctgactg
1251 accgcgttac tcccacaggt gagcggggcg gacggccctt ctctccggg
1301 ctgtaattag cgcttggttt aatgacggct tgtttctttt ctgtggctgc
1351 gtgaaagcct tgaggggctc cgggagggcc ctttgtgcgg gggggagcgg
1401 ctcggggggt gcgtgcgtgt gtgtgtgcgt ggggagcgcc gcgtgcggcc
1451 cgcgtgccc gcgggtgtg agcgtgcgg gcgcggcgcg gggctttgtg
1501 cgctccgcag tgtgcgcgag gggagcgcg ccggggcgcg tgcccccgcg
1551 tgcggggggg gctgcgaggg gaacaaaggc tgcgtgcggg gtgtgtgcgt
1601 ggggggggtg gcaggggggt tgggcgcggc ggtcgggctg taaccccccc
1651 ctgcaccccc ctcccagagt tgctgagcac ggcccggctt cgggtgcggg
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1751 ggcaggtggg ggtgccgggc gggcgggggc gcctcgggc cggggagggc
1801 tcgggggagg ggcgcggcg ccccgaggc gccggcggtc gtcgagcgcg
1851 ggcgagccgc agccattgcc ttttatggta atcgtgcgag agggcgagg
1901 gacttccttt gtcccaaate tgtgcggagc cgaaatctgg gaggcgccgc
1951 cgcacccccct ctagcgggcg cggggcgaa ggtgcggcg ccggcaggaa
2001 ggaaatgggc ggggaggggc ttcgtgcgtc gccgcgccgc cgtccccttc
2051 tccctctcca gcctcggggc tgtccgcggg gggacggctg cttcggggg
2101 ggacggggca gggcggggtt cggcttcttg cgtgtgaccg gcggtctag
2151 agcctctgct aaccatgttc atgccttctt ctttttcta cagctcctgg
2201 gcaacgtgct ggttattgtg ctgtctcatc attttggcaa agaattcgat
2251 atcaagcttg gggattttca ggcaccacca ctgacctggg acagtgaatc
2301 gacaatgccg tcttctgtct cgtggggcat cctcctgctg gcaggcctgt
2351 gctgcctggg ccctgtctcc ctggctgagg atccccagg agatgctgcc
2401 cagaagacag atacatccca ccatgatcag gatcacccaa ccttcaacaa
```

FIGURE 24A

p43msENCB-AT

Page 2

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2451 gatcaccccc aacctggctg agttcgcctt cagcctatac cgccagctgg
2501 cacaccagtc caacagcacc aatatcttct tctccccagt gagcatcgct
2551 acagcctttg caatgctctc cctggggacc aaggctgaca ctcacgatga
2601 aatcctggag ggcctgaatt tcaacctcac ggagattccg gaggtcaga
2651 tccatgaagg cttccaggaa ctctccgta ccctcaacca gccagacagc
2701 cagctccagc tgaccaccgg caatggcctg ttctcagcg agggcctgaa
2751 gctagtggat aagtttttgg aggatgttaa aaagtgtgac cactcagaag
2801 ccttactgt caacttcggg gacaccgaag aggccaagaa acagatcaac
2851 gattacgtgg agaagggtac tcaagggaaa attgtggatt tgggtcaagga
2901 gcttgacaga gacacagttt ttgctctggt gaattacatc ttctttaaag
2951 gcaaattgga gagacccttt gaagtcaagg acaccgagga agaggacttc
3001 cacgtggacc aggtgaccac cgtgaagggt cctatgatga agcgtttagg
3051 catgttttaac atccagcact gtaagaagct gtccagctgg gtgctgctga
3101 tgaaatacct gggcaatgcc accgccatct tcttcttgcc tgatgagggg
3151 aaactacagc acctggaaaa tgaactcacc cagcatatca tcaccaagtt
3201 cctggaaaat gaagacagaa ggtctgccag cttacattta cccaaactgt
3251 ccattactgg aacctatgat ctgaagagcg tcctgggtca actgggcatc
3301 actaaggtct tcagcaatgg ggctgacctc tccgggggtca cagaggaggc
3351 acccctgaag ctctccaagg ccgtgcataa ggctgtgctg accatcgacg
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3551 cccaaaaata actgcctctc gctcctcaac ccctccctc catccctggc
3601 cccctccctg gatgacatta aagaagggtt gagctggtaa ccccccccc
3651 ccctgcaggg gccctcgacc cgggcgccg cttcgagcag acatgataag
3701 atacattgat gagtttgac aaaccacaac tagaatgcag tgaaaaaaat
3751 gctttatttg tgaaatttgt gatgctattg ctttatttgt aaccattata
3801 agctgcaata aacaagttaa caacaacaat tgcattcatt ttatgtttca
3851 ggttcagggg gagatgtggg aggtttttta aagcaagtaa aacctctaca
3901 aatgtggtaa aatcgataag gatctaggaa ccctagtga tggagttggc
3951 cactccctct ctgcgcgctc gctcgctcac tgaggccgcc cgggcaaagc
4001 ccgggcgctc ggcgaccttt ggtcgcccg cctcagtgag cgagcgagcg
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4101 taatagcgaa gagggccgca ccgatcgccc ttcccaacag ttgcgtagcc
4151 tgaatggcga atggcgcgac gcgcctgta gcggcgcat aagcgcgcg
4201 ggtgtggtgg ttacgcgcag cgtgaccgt acacttgcca gcgccttagc
4251 gcccgctcct ttcgctttct tcccttcctt tctcgccacg ttcgcccgt
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4351 gctttacggc acctcgacct caaaaaactt gattaggggt atggttcacg
4401 tagtgggcca tcgccctgat agacggtttt tcgcccttg acgttgaggt
4451 ccacgttctt taatagtgga ctcttgttcc aaactggaac aacactcaac
4501 cctatctcgg tctattcttt tgatttataa gggattttgc cgatttcggc
4551 ctattggtta aaaaatgagc tgatttaaca aaaatttaac gcgaatttta
4601 acaaaatatt aacgtttaca atttcctgat gcggtatttt ctccttacgc
4651 atctgtgcgg tatttcacac cgcatatggt gcactctcag tacaatctgc
4701 tctgatgccg catagttaag ccagccccga caccgcgcaa caccgcgtga
4751 cgcgccctga cgggcttgct tgcctccggc atccgcttac agacaagctg
4801 tgaccgtctc cgggagctgc atgtgtcaga ggttttcacc gtcacaccg
4851 aaacgcgcga gacgaaaggg cctcgtgata cgcctatttt tataggttaa
4901 tgtcatgata ataattggttt cttagacgtc aggtggcact tttcggggaa
4951 atgtgcgcgg aaccctatt tgtttatttt tctaaataca ttcaaatatg
5001 tatccgctca tgagacaata accctgataa atgcttcaat aatattgaaa
5051 aaggaagagt atgagtattc aacatttcgg tgctgccttt attccctttt
5101 ttgcggcatt ttgccttctt gtttttgctc acccagaaac gctgtgaaa
5151 gtaaaagatg ctgaagatca gttgggtgca cgagtgggtt acatcgaaact

```

FIGURE 24B

p43msENCB-AT

Page 3

5201	ggatctcaac	agcggtaaga	tccttgagag	ttttcgcccc	gaagaacggt
5251	ttccaatgat	gagcactttt	aaagtctctgc	tatgtggcgc	ggtattatcc
5301	cgtattgacg	ccgggcaaga	gcaactcggg	cgccgcatac	actattctca
5351	gaatgacttg	gttgagtact	caccagtcac	agaaaagcat	cttacggatg
5401	gcatgacagt	aagagaatta	tgcagtgtcg	ccataaccat	gagtataaac
5451	actgcgccca	acttacttct	gacaacgatc	ggaggaccga	aggagctaac
5501	cgcttttttg	cacaacatgg	gggatcatgt	aactcgcctt	gatcgttggg
5551	aaccggagct	gaatgaagcc	ataccaaacg	acgagcgtga	caccacgatg
5601	cctgtagcaa	tggcaacaac	gttgcgcaaa	ctattaactg	gcgaactact
5651	tactctagct	tcccggcaac	aattaataga	ctggatggag	gcgataaaag
5701	ttgcaggacc	acttctgcgc	tcggcccttc	cggtctggctg	gtttattgct
5751	gataaatctg	gagccgggtga	gcgtgggtct	cgcggtatca	ttgcagcact
5801	ggggccagat	ggtaagccct	cccgtatcgt	agttatctac	acgacgggga
5851	gtcaggcaac	tatggatgaa	cgaaatagac	agatcgctga	gatagggtcc
5901	tcactgatta	agcattggta	actgtcagac	caagtttact	catatatact
5951	ttagattgat	ttaaaaacttc	atttttaatt	taaaaggatc	taggtgaaga
6001	tcctttttga	taatctcatg	acaaaaatcc	cttaacgtga	gttttcgttc
6051	cactgagcgt	cagaccccgt	agaaaagatc	aaaggatcct	cttgagatcc
6101	tttttttctg	cgcgtaatct	gctgcttgca	aacaaaaaaa	ccaccgctac
6151	cagcggtggt	ttgtttgccg	gatcaagagc	taccaactct	ttttccgaag
6201	gtaactggct	tcagcagagc	gcagatacca	aatactgtcc	ttctagtgtg
6251	gccgtagtta	ggccaccact	tcaagaactc	tgtagcaccg	cctacatacc
6301	tcgctctgct	aatcctgtta	ccagtggctg	ctgccagtgg	cgataagtcg
6351	tgtcttaccg	ggttggactc	aagacgatag	ttaccggata	aggcgcagcg
6401	gtcgggctga	acgggggggtt	cgtgcacaca	gccagcttg	gagcgaacga
6451	cctacaccga	actgagatac	ctacagcgtg	agcattgaga	aagcgccacg
6501	cttcccgaag	ggagaaaaggc	ggacaggtat	ccggtaagcg	gcagggtcgg
6551	aacaggagag	cgcacgaggg	agcttccagg	gggaaacgcc	tggatatctt
6601	atagtcctgt	cgggtttcgc	cacctctgac	ttgagcgtcg	atttttgtga
6651	tgctcgtcag	gggggcggag	cctatggaaa	aacgccagca	acgcggcctt
6701	tttacggttc	ctggcctttt	gctggccttt	tgctcacatg	ttctttcctg
6751	cgttatcccc	tgattctgtg	gataaccgta	ttaccgcctt	tgagtgaact
6801	gataccgctc	gccgcagccg	aacgaccgag	cgcagcgagt	cagtgagcga
6851	ggaagcggaa	gagcgcccaa	tacgcaaacc	gcctctcccc	gcgcgttggc
6901	cgattcatta	atgcagggct	gcag		

FIGURE 24C

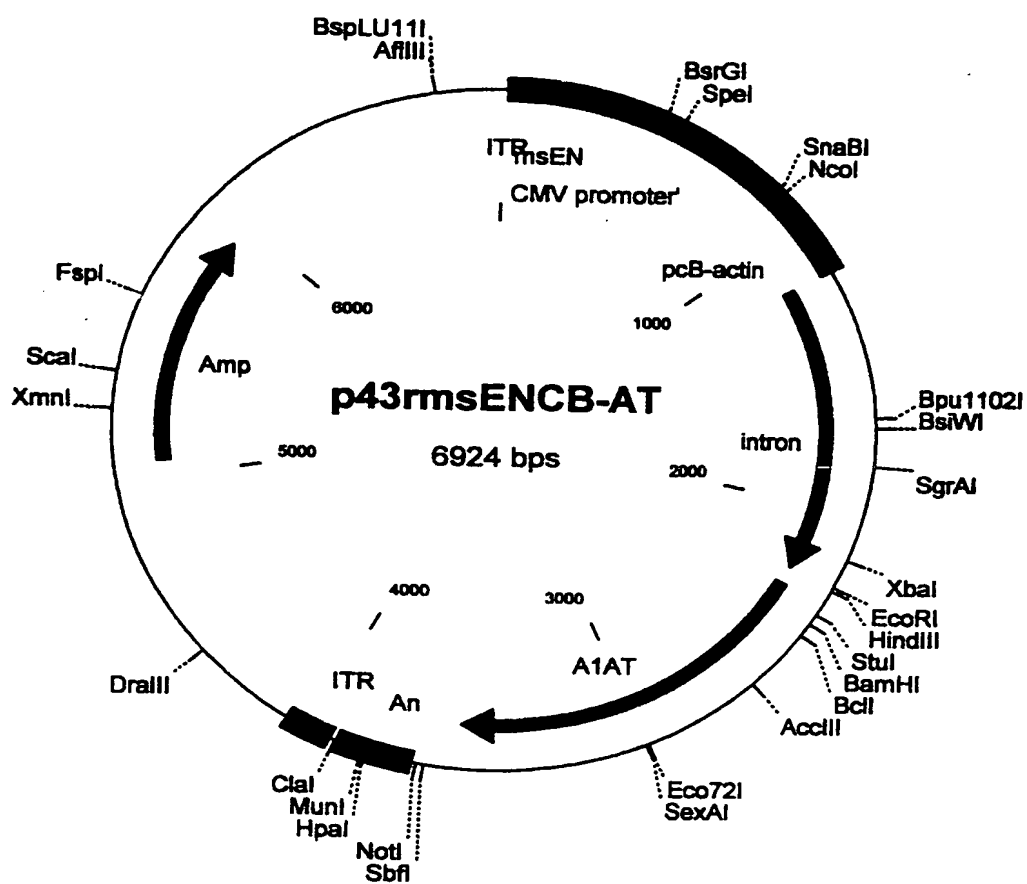


FIGURE 25

19 Apr 1999

Sequence Data

Page 1

Molecule: p43rmsENCB-AT, 6924 bps DNA Circular
Description: Ligation of inverted msEnhacer into p43CB-AT*
File Name: p43rmsCB-AT.cm5, dated 19 Apr 1999
Printed: 1-6924 bps (Full), format Single Strand

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51  actgaggccg ggcgaccaaa ggtcgcccga cgcccgggct ttgcccgggc
101 ggcctcagtg agcgagcgag cgcgagagaga gggagtggcc aactccatca
151 ctaggggttc ctatgtctga cacccaaata tggcctgggg tgaggaatgg
201 tgccgtcgcc atatttgggt gtccaccatt cctcaccgct ctaaaaataa
251 ctcccgggag ttatttttag agcgccaaca cctgctgcct gccaccatt
301 cctcaccgct ctaaaaataa ctccccacca ttcctcaccg gtcgccatat
351 ttgggtgtcg tgaggaatgg tgagatcttc aatattggcc attagccata
401 ttattcattg gttatatagc ataaatcaat attggctatt ggccattgca
451 tacgttgtat ctatatcata atatgtacat ttatatggc tcattgtccaa
501 tatgaccgcc atgttggcat tgattattga ctagtattta atagtaatca
551 attacggggt cattagttca tagcccatat atggagtacc gcgttacata
601 acttacggta aatggcccgc ctggctgacc gcccaacgac ccccgcccat
651 tgacgtcaat aatgacgtat gtcccataag taacgccaat agggactttc
701 cattgacgtc aatgggtgga gtatttacgg taaactgccc acttggcagt
751 acatcaagtg tatcatatgc caagtccgcc ccctattgac gtcaatgacg
801 gtaaatggcc cgcttggcat tatgcccagt acatgacctt acgggacttt
851 cctacttggc agtacatcta cgtattagtc atcgctatta ccatggtcga
901 ggtgagcccc acgttctgct tcactctccc catctcccc cctccccac
951 cccaatttt gtatttattt atttttaaat tattttgtgc agcgatgggg
1001 gcgggggggg ggggggggag cgcgccaggc ggggcggggc ggggcgaggc
1051 gcggggaggg ggcgagcgga gaggtgcggc ggcagccaat cagagcgggc
1101 cgctccgaaa gtttcctttt atggcgaggc ggcggcgggc gcgggccctat
1151 aaaaagcgaa gcgcgcggcg ggcgggagtc gctgcgacgc tgccttcgcc
1201 ccgtgccccg ctccgcccgc gcctcgcgcc gcccgccccg gctctgactg
1251 accgcgttac tcccacaggt gagcgggcgg gacggccctt ctccctccggg
1301 ctgtaattag cgcttggttt aatgacggct tgtttctttt ctgttgctgc
1351 gtgaaagcct tgaggggctc cgggagggcc ctttgtgcgg gggggagcgg
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1501 cgctccgcag tgtgcgcgag ggaagcgcgg ccggggggcg tgccccgcgg
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1651 ctgcaccccc ctccccgagt tgctgagcac ggcccgggtt cgggtgcggg
1701 gctccgtacg gggcgtggcg cggggctcgc cgtgccgggc ggggggtggc
1751 ggcaggtggg ggtgccgggc ggggcggggc cgcctcgggc cggggagggc
1801 tcgggggagg ggcgcggcgg ccccggagc gccggcggtt gtcgagggcg
1851 ggcgagccgc agccattgcc ttttatggta atcgtcgagc agggcgaggg
1901 gacttccttt gtcccaaate tgtgcggagc cgaaatctgg gaggcgccgc
1951 cgcacccccc cttagcgggag cggggcgaag cgggtgcggc ccggcaggaa
2001 ggaaatgggc ggggagggcc ttcgtgcgtc gccgcgccgc cgtccccttc
2051 tccctctcca gcctcggggc tgtccgcggg gggacggctg ccttcggggg
2101 ggacggggca gggcgggggt atgcttcttg cgtgtgaccg gcggtcttag
2151 agcctctgct aaccatgttc atgccttctt ctttttctta cagtctctgg
2201 gcaacgtgct ggttattgtg ctgtctcatc attttggcaa agaattcgat
2251 atcaagcttg gggattttca ggcaccacca ctgacctggg acagtgaatc
2301 gacaatgccg tcttctgtct cgtggggcat cctcctgctg gcaggcctgt
2351 gctgcctggt cctgtctccc ctggctgagg atccccagg agatgctgcc
2401 cagaagacag atacatccca ccatgatcag gatcacccaa ccttcaacaa
```

FIGURE 25A

p43rmsENCB-AT

Page 2

2451	gatcaccccc	aacctggctg	agttcgcctt	cagcctatac	cgccagctgg
2501	cacaccagtc	caacagcacc	aatatcttct	tctccccagt	gagcatcgct
2551	acagcctttg	caatgctctc	cctggggacc	aaggctgaca	ctcacgatga
2601	aatcctggag	ggcctgaatt	tcaacctcac	ggagattccg	gaggctcaga
2651	tccatgaagg	cttccaggaa	ctcctccgta	ccctcaacca	gccagacagc
2701	cagctccagc	tgaccaccgg	caatggcctg	ttcctcagcg	agggcctgaa
2751	gctagtggat	aagtttttgg	aggatgttaa	aaagttgtac	cactcagaag
2801	ccttcactgt	caacttcggg	gacaccgaag	aggccaagaa	acagatcaac
2851	gattacgtgg	agaagggtac	tcaagggaaa	attgtggatt	tggtcaagga
2901	gcttgacaga	gacacagttt	ttgctctggt	gaattacatc	ttctttaaag
2951	gcaaatggga	gagacccttt	gaagtcaagg	acaccgagga	agaggacttc
3001	cacgtggacc	aggtgaccac	cgtgaagggtg	cctatgatga	agcgtttagg
3051	catgtttaac	atccagcact	gtaagaagct	gtccagctgg	gtgctgctga
3101	tgaaatacct	gggcaatgcc	accgccatct	tcttcctgcc	tgatgagggg
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3201	cctggaaaat	gaagacagaa	ggtctgccag	cttacattta	cccaaactgt
3251	ccattactgg	aacctatgat	ctgaagagcg	tcctgggtca	actgggcatac
3301	actaaggtct	tcagcaatgg	ggctgacctc	tcgggggtca	cagaggaggc
3351	acccctgaag	ctctccaagg	ccgtgcataa	ggctgtgctg	accatcgacg
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3451	tctatccccc	ccgaggtcaa	gttcaacaaa	ccctttgtct	tcttaatgat
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3551	cccaaaaata	actgcctctc	gctcctcaac	ccctcccctc	catccctggc
3601	ccctcccttg	gatgacatta	aagaagggtt	gagctggtaa	ccccccccc
3651	ccctgcaggg	gccctcgacc	cgggcgcccg	cttcgagcag	acatgataag
3701	atacattgat	gagtttggac	aaaccacaac	tagaatgcag	tgaaaaaaat
3751	gctttatttg	tgaaatttgt	gatgtatttg	ctttatttgt	aaccattata
3801	agctgcaata	aacaagttaa	caacaacaat	tgcatctcatt	ttatgtttca
3851	ggttcagggg	gagatgtggg	aggtttttta	aagcaagtaa	aacctctaca
3901	aatgtggtaa	aatcgataag	gatctaggaa	cccctagtga	tggagttggc
3951	cactccctct	ctgcgcgctc	gctcgcctcac	tgaggccgcc	cgggcaaaagc
4001	ccgggcgctc	ggcgaccttc	ggtcgcccg	cctcagtgag	cgagcgagcg
4051	cgagagaggg	gagtggccaa	ccccccccc	ccccccctg	cagcctggcg
4101	taatagcgaa	gaggcccgc	ccgatcgccc	ttcccaacag	ttgctgtagc
4151	tgaatggcga	atggcgcgac	gcgcctgtga	gcggcgcat	aagcgcgcg
4201	ggtgtggtgg	ttacgcgcag	cgtgaccgct	acacttgcca	gcgccttagc
4251	gcccgcctct	ttcgctttct	tcccttcctt	tctcgccacg	ttcgccggct
4301	ttccccgtca	agctctaaat	cgggggctcc	ctttagggtt	ccgatttagt
4351	gctttacggc	acctcgaccc	caaaaaactt	gattagggtg	atggttcacg
4401	tagtgggcca	tcgcccctgat	agacgggttt	tcgccccttg	acgttggagt
4451	ccacgttctt	taatagtgg	ctcttggtcc	aaactggaac	aacactcaac
4501	cctatctcgg	tctattcttt	tgatttataa	gggattttgc	cgatttcggc
4551	ctattgggtta	aaaaatgagc	tgattttaaca	aaaattttaac	gcgaatttta
4601	acaaaatatt	aacgttttaca	atttcctgat	gcggtatttt	ctccttacgc
4651	atctgtgcgg	tatttcacac	cgcatatggt	gcactctcag	tacaatctgc
4701	tctgatgccg	catagttaa	ccagccccga	caccgcgcaa	caccgctga
4751	cgcgccctga	cggtcttgct	tgctcccggc	atccgcttac	agacaagctg
4801	tgaccgtctc	cgggagctgc	atgtgtcaga	ggttttcacc	gtcatcaccg
4851	aaacgcgcga	gacgaaaggg	cctcgtgata	cgctattttt	tatagggttaa
4901	tgtcatgata	ataatgggtt	cttagacgtc	aggtggcact	tttcggggaa
4951	atgtgcgcgg	aacccttatt	tgttttattt	tctaaataca	ttcaaatatg
5001	tatccgctca	tgagacaata	accctgataa	atgcttcaat	aatattgaaa
5051	aaggaagagt	atgagtattc	aacatttccg	tgctcgccctt	attccctttt
5101	ttgcggcatt	ttgccttcct	gtttttgctc	accagaaaac	gctggtgaaa
5151	gtaaaagatg	ctgaagatca	gttggggtgca	cgagtgggtt	acatcgaaat

FIGURE 25B

p43rmsENCB-AT

Page 3

5201	ggatctcaac	agcggtaaga	tccttgagag	ttttcgcccc	gaagaacggt
5251	ttccaatgat	gagcactttt	aaagttctgc	tatgtggcgc	ggtattatcc
5301	cgtattgacg	ccgggcaaga	gcaactcggg	cgccgcatac	actattctca
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5451	actgcggcca	acttacttct	gacaacgac	ggaggaccga	aggagctaac
5501	cgcttttttg	cacaacatgg	gggatcatgt	aactcgcctt	gatcggtggg
5551	aaccggagct	gaatgaagcc	ataccaaacg	acgagcgtga	caccacgatg
5601	cctgtagcaa	tggcaacaac	gttgcgcaaa	ctattaactg	gcgaactact
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5801	ggggccagat	ggtaagccct	cccgtatcgt	agttatctac	acgacgggga
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5901	tcactgatta	agcattggta	actgtcagac	caagtttact	catatatact
5951	ttagattgat	ttaaaacttc	atttttaatt	taaaaggatc	taggtgaaga
6001	tcctttttga	taatctcatg	acaaaaatcc	cttaacgtga	gttttcgttc
6051	cactgagcgt	cagaccccg	agaaaagatc	aaaggatctt	cttgagatcc
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6151	cagcggtggt	ttgtttgccc	gatcaagagc	taccaactct	ttttccgaag
6201	gtaactggct	tcagcagagc	gcagatacca	aatactgtcc	ttctagtgtg
6251	gccgtagtta	ggccaccact	tcaagaactc	tgtagcaccg	cctacatacc
6301	tcgctctgct	aatcctgtta	ccagtggctg	ctgccagtgg	cgataagtgc
6351	tgtcttaccg	ggttggactc	aagacgatag	ttaccggata	aggcgcagcg
6401	gtcgggctga	acgggggggt	cgtgcacaca	gcccagcttg	gagcgaacga
6451	cctacaccga	actgagatac	ctacagcgtg	agcattgaga	aagcgccacg
6501	cttcccgaag	ggagaaaaggc	ggacaggat	ccggtaagcg	gcagggtcgg
6551	aacaggagag	cgcacgaggg	agcttccagg	gggaaacgcc	tggtatcttt
6601	atagtcctgt	cggttttcgc	cacctctgac	ttgagcgtcg	atttttgtga
6651	tgctcgtcag	gggggcggag	cctatggaaa	aacgccagca	acgcggcctt
6701	tttacggttc	ctggcctttt	gctggccttt	tgctcacatg	ttctttcctg
6751	cgttatcccc	tgattctgtg	gataaccgta	ttaccgcctt	tgagtgcgct
6801	gataaccgctc	gccgcagccg	aacgaccgag	cgcagcgagt	cagtgcgcga
6851	ggaagcggaa	gagcgcccaa	tacgcaaacc	gcctctcccc	gcgcgttggc
6901	cgattcatta	atgcagggct	gcag		

FIGURE 25C

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